ACTIVE MICRO TECHNOLOGIES

Technical Dossier

FortiCulture Coconut

Code Number: M11001

INCI Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate

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FortiCulture Coconut

Technical Data Sheet

BACKGROUND

With the rise of gut health and importance of maintaining the body's immune system, consumers are searching for solutions that take their holistic health into consideration in the form of immunocosmetics that protect the microbiome. Skin is the body's fortress. However, external stressors like pollution, UV radiation, and pathogenic bacteria attempt to break down and disrupt the barrier daily. The key to the microbiome lies in understanding how we can use our biology to guard ourselves from damage.

After the successful launch of PhytoCide Lichen for anionic systems, many formulators and brands alike demanded yet another formulation friendly antimicrobial, except this time... with probiotic provenance to address the growing interest in the skin's microbiome. This challenged us like never before to combine our years of expertise in natural antimicrobials with immunocosmetics, and develop **FortiCulture Coconut** to deliver a non-irritating, effective, and multifunctional product capable of providing broad-spectrum protection to formulations and fortifying the skin barrier for anti-aging benefits.

SCIENCE

Inspired by previous research done by the University of California Davis investigating the novel medical applications of antimicrobial peptides (AMPs), Active Micro Technologies realized they showed exceptional promise in personal care applications. After all, AMPs are more than just simple peptides, they are signaling molecules that are critical for maintaining skin barrier homeostasis. Our skin naturally creates its own AMPs if toll-like receptors (TLRs) sense a particular signaling pattern. Traditionally, TLRs required some source of inflammation to trigger the release of the AMPs, but not anymore. When topically applied to the skin, **FortiCulture Coconut**'s standardized lipopeptide content helps mimic the cellular signaling pattern which encourages TLRs to release AMPs, without the need for inflammation. When AMPs are released, this instructs our skin's immune cells to boost the skin barrier through keratinization which increases collagen and reduces signs of cellular aging. **FortiCulture Coconut** emerges as the lipopeptide-powered natural antimicrobial that fortifies the skin and formulations from the inside-out.

Code: M11001
INCI: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate
INCI Status: Conforms
REACH Status: Compliant
CAS: 7732-18-5 & 8001-31-8 & 8013-01-2
EINECS: 231-791-2 & 232-282-8 & 232-387-9
Origin: Botanical
Processing:
- GMO Free
- No Ethoxylation
- No Irradiation
- No Sulphonation
- No Ethylene Oxide treatment
- No Hydrogenation

Additives: None
- Preservatives: None
- Antioxidants: None

Solvents used: Water
Appearance: Clear to Slightly Hazy Liquid
Soluble/Miscible: Water Soluble
Suggested Use Levels: 1.0 - 4.0%
Suggested Applications:
Barrier Boosting, Antimicrobial, Anti-aging

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Version 1 - 03-08-2024

FortiCulture Coconut

BENEFITS

By combining our Cradle to Cradle sustainability approach with our fermentation expertise, Active Micro Technologies introduces **FortiCulture Coconut**: A lipopeptide-powered natural antimicrobial which activates cellular signaling to elicit anti-aging benefits from within. **FortiCulture Coconut** instructs keratinocytes to synthesize antimicrobial peptides to fortify the skin barrier and encourage homeostasis.

To verify this, SDS Gel Electrophoresis is employed as a qualitative and quantitative method to measure protein molecular weight using polyacrylamide gel and electrolysis. Human epidermal keratinocytes were incubated in complete media and increasing concentrations of **FortiCulture Coconut**. The migration rate of bands directly correlates with the keratinocyte molecular weight in Figure 1. Equal areas of interest are identified, and are further analyzed using ImageJ histogram software. The difference in luminosity can be used to measure the amount of protein content. More protein is a darker blue, and the histogram output will read as a left-leaning, lower intensity in Figure 2. **FortiCulture Coconut** at 0.04% increased keratinocyte molecular weight by 13%, indicating the synthesis of antimicrobial peptides.

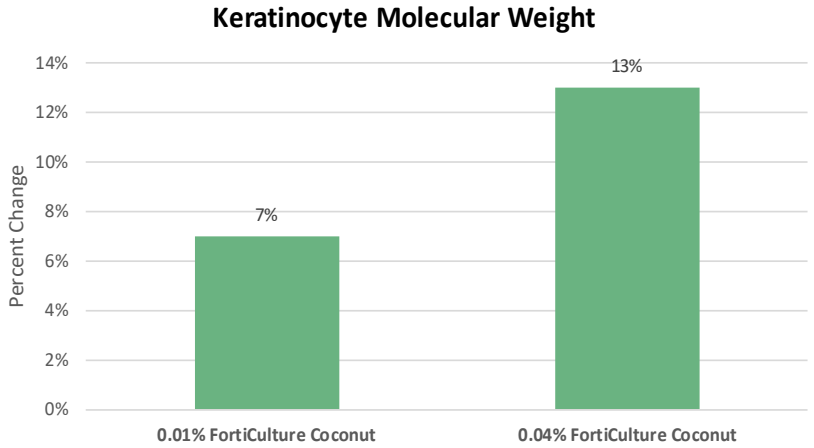

Figure 1. Percent Change in Keratinocyte Molecular Weight Compared to Complete Media.

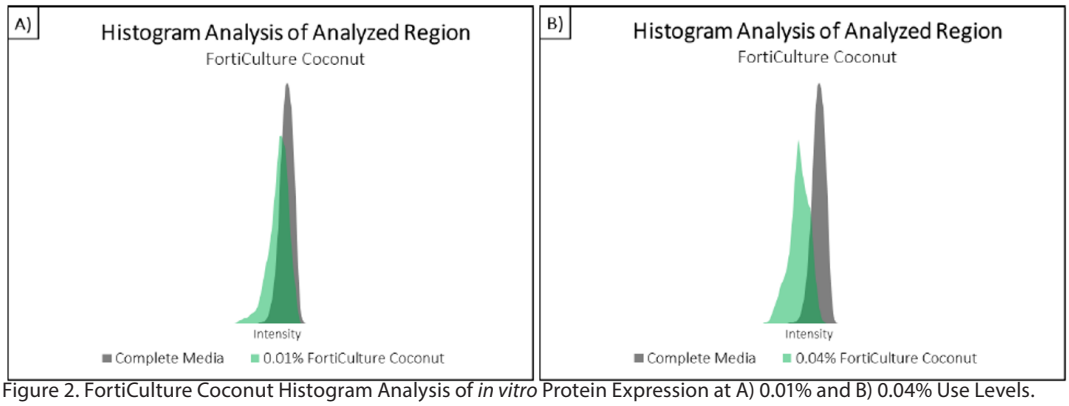

Figure 2. FortiCulture Coconut Histogram Analysis of *in vitro* Protein Expression at A) 0.01% and B) 0.04% Use Levels.

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FortiCulture Coconut

Microorganism Tested	MIC (%)
E. coli	0.50
P. aeruginosa	0.50
S. aureus	1.00
C. albicans	1.00
A. brasiliensis	1.00

Figure 3. MIC Data for **FortiCulture Coconut**

One of the first steps in the development of this product was to determine the potential ability to inhibit the growth of a variety of bacteria and fungi. Using standard serial dilution protocols in growth media, the Minimum Inhibitory Concentrations (MICs) for **FortiCulture Coconut** were determined for a variety of both bacterial and fungal organisms. The results of these tests are shown in Figure 3.

The positive MIC screening results warranted further testing to confirm its ability to provide product preservation. A Double Challenge Test was completed using 1.0% **FortiCulture Coconut** in the water phase of a generic cream base formulation. Samples were inoculated with *E. coli, P. aeruginosa*, *S. aureus*, *C. albicans*, and *A. brasiliensis*. During the first 28-day incubation period, samples were periodically collected and tested for the presence of these microorganisms. Following this initial 28 days of incubation, the cream samples were then re-inoculated with the microbial cultures and sampled over an additional 28-day period. Figure 4 shows the positive preservation results for 1.0% **FortiCulture Coconut**.

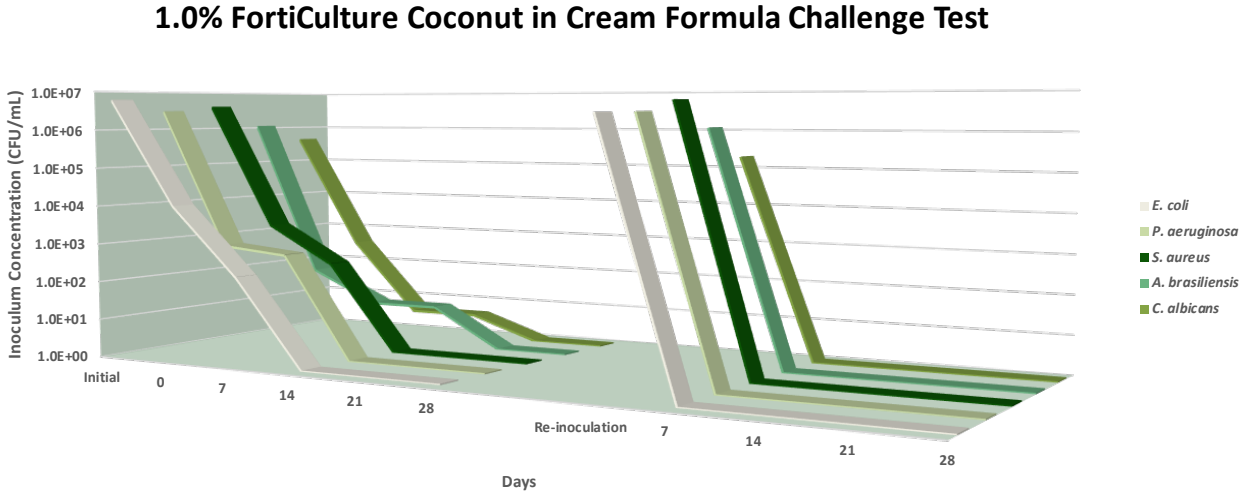

Figure 4. Challenge Test results for Generic Cream Formula with 1.0% **FortiCulture Coconut** inoculated on Day 0 and re-inoculated on Day 28. Results show log reduction in viable organisms.

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A ROS Scavenging Assay was conducted to assess the *in vitro* effect of **FortiCulture Coconut** to scavenge unnecessary oxidative stress in dermal fibroblasts. Attenuating excessive ROS preserves cellular homeostasis and blunts intrinsic and extrinsic age-related declines in skin cell function. Compared to untreated fibroblasts, AntA (200 pM) increased ROS levels as expected, indicating a valid test. Fibroblasts treated with **FortiCulture Coconut** at 0.01%, 0.1%, and 1.0% exhibited a reduction in oxidative stress compared to those exposed to AntA, pictured in Figure 4.

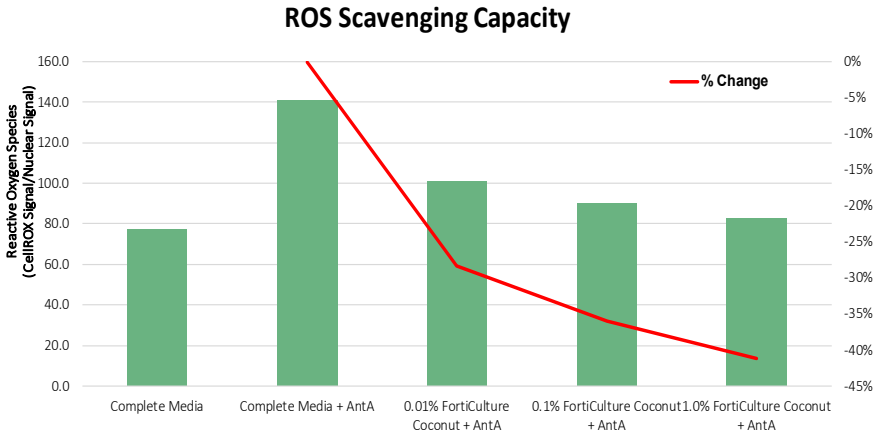

Figure 4. The effect of **FortiCulture Coconut** on ROS scavenging.

A cellular aging model was developed to assess the *in vitro* effect of **FortiCulture Coconut** to reduce SA-Beta-gal activity in "aged" fibroblasts. Cellular senescence is a state of permanent cell cycle arrest that accompanies aging and contributes to a decline in normal skin function and physiology. SA-Beta-gal is the gold standard biomarker to identify senescence *in vitro* as the enzyme, beta-galactosidase, explicitly accumulates in the lysosomes of senescent cells. **FortiCulture Coconut** at 0.01% and 0.1% concentrations elicited a 25% and 28% reduction in SA-Beta-gal Activity compared to untreated fibroblasts, as depicted in Figure 5. **FortiCulture Coconut** reduces cellular senescence and may attenuate or reverse the alterations in skin structure and physiology that occur during aging.

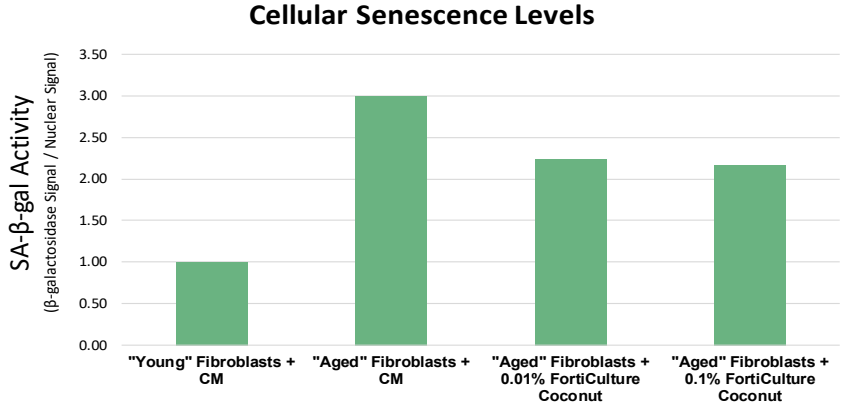

Figure 5. The effect of **FortiCulture Coconut** on cellular senescence.

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Finally, a Sirius Red/Fast Green Collagen Assay was conducted to assess the *in vitro* effect of **FortiCulture Coconut** to trigger collagen synthesis in dermal fibroblasts. Human dermal fibroblasts were seeded into 24-well tissue culture plates and allowed to grow to confluency in complete media (CM). 0.01%, 0.02%, and 0.04% concentrations of **FortiCulture Coconut** were diluted in serum-free CM and incubated with fibroblasts for 24 hours. AA2G (100 µM) and IGF-1 (50 ng/mL) were diluted in serum-free CM and utilized as positive controls. Results depicted in Figure 6 indicate that **FortiCulture Coconut** stimulates collagen synthesis which may lead to improvements in dermal-epidermal junction integrity and provide a healthier complexion.

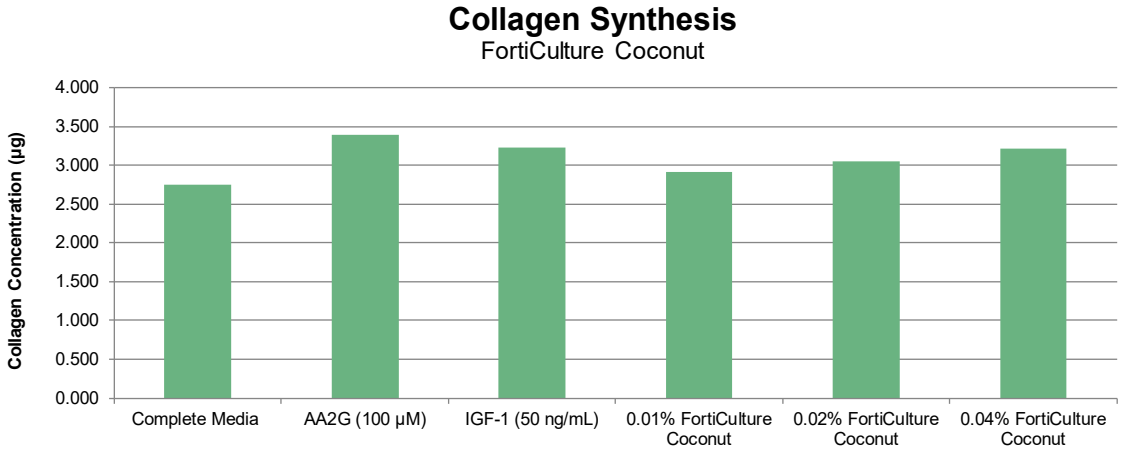

Figure 6. The effect of **FortiCulture Coconut** on collagen synthesis.

USE RECOMMENDATIONS

FortiCulture Coconut is water-soluble, and can be incorporated into aqueous systems, as well as the aqueous phase of emulsions. When using **FortiCulture Coconut** it is recommended to maintain the formulation pH between 3 and 8. We also suggest incorporating the product at or below 40°C. **FortiCulture Coconut** is non-ionic in nature, and has excellent compatibility with both cationic and anionic systems.

REFERENCES

1. ADM Outside Voice Research. "ADM Identifies 2022 Global Consumer Trends" 2022.
2. Cui CY, Schlessinger D. Eccrine sweat gland development and sweat secretion. Exp Dermatol. 2015;24(9):644–650.
3. Clausen ML, Agner T. Antimicrobial peptides, infections and the skin barrier. Curr Probl Dermatol. 2016;49:38–46.
4. Sørensen OE, Cowland JB, Theilgaard-Mönch K, Liu L, Ganz T, Borregaard N. Wound healing and expression of antimicrobial peptides/polypeptides in human keratinocytes, a consequence of common growth factors. J Immunol. 2003;170(11):5583–5589.

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Specification

Product Name: FortiCulture Coconut
Code Number: M11001
CAS #'s: 7732-18-5 & 8001-31-8 & 8013-01-2
EINECS #'s: 231-791-2 & 232-282-8 & 232-387-9
INCI Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate

Specification	Parameter
Appearance	Clear to Slightly Hazy Liquid
Color	Colorless to Light Yellow
Odor	Characteristic
pH (Direct)	3.8 – 5.5
NVM (1g-1hr-105°C)	16.0 – 22.0%
Lipopeptide Content (Kjeldahl Nitrogen x 6.25)	15.0 – 20.0%
Keratinocyte Expressed Proteins (KEP) > 5%	Positive
Heavy Metals Lead Arsenic Cadmium	< 20 ppm < 10 ppm < 2 ppm < 1 ppm
Minimum Inhibitory Concentration[1] Organism (ATCC#)	
E. coli (#8739)	0.50 – 2.00%
S. aureus (#6538)	1.00 – 2.00%
P. aeruginosa (#9027)	0.25 – 1.00%
C. albicans (#10231)	1.00 – 2.00%
A. brasiliensis (#16404)	1.00 – 2.00%

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DO NOT FREEZE; Store at or near room temperature;
May sediment upon standing; Mix well prior to use

Product may change appearance if exposed to cold temperatures during shipment or storage. If this happens, please gently warm to 45-50°C and mix until normal appearance is restored.

Note:

1) Refer to Inhibition Activity Data

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Compositional Breakdown

FortiCulture Coconut
Code: M11001

Compositional Breakdown:

Ingredient	%
Water	80.00
Cocos Nucifera (Coconut) Fruit Extract	10.00
Saccharomyces Ferment Filtrate	10.00

Compositional Breakdown

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Active Micro Technologies hereby confirms that to the best of our knowledge, none of the potential 26 fragrance allergens listed below are present in our finished product or as an intentional component in the raw materials used to manufacture this product. We do not routinely analyze our product for the substances listed below:

ALLERGENS listed in Annex III of EU Cosmetic Regulation(EC) No. 1223/2009	
INCI NAME	CAS Number
Alpha-Isomethyl Ionone	127-51-5
Amyl Cinnamal	122-40-7
Amylcinnamyl Alcohol	101-85-9
Anise Alcohol	105-13-5
Benzyl Alcohol	100-51-6
Benzyl Benzoate	120-51-4
Benzyl Cinnamate	103-41-3
Benzyl Salicylate	118-58-1
Butylphenyl Methylpropional	80-54-6
Cinnamal	104-55-2
Cinnamyl Alcohol	104-54-1
Citral	5392-40-5
Citronellol	106-22-9
Coumarin	91-64-5
Eugenol	97-53-0
Evernia Furfuracea (Treemoss) Extract	90028-67-4
Evernia Prunastri (Oakmoss) Extract	90028-68-5
Farnesol	4602-84-0
Geraniol	106-24-1
Hexyl Cinnamal	101-86-0
Hydroxycitronellal	107-75-5
Hydroxyisohexyl 3-Cyclohexene Carboxaldehyde (Lyral)	31906-04-4
Isoeugenol	97-54-1
Limonene (sum of d, l and dl)	5989-27-5
Linalool	78-70-6
Methyl 2-Octynoate	111-12-6

Compositional Breakdown

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Active Micro Technologies hereby confirms that to the best of our knowledge, none of the pesticides listed below are present in our finished product or as an intentional component in the raw material used to manufacture this product. We do not routinely analyze our product for the substances listed below:

INCI NAME	CAS Number
Alachlor	15972-60-8
Aldrin	309-00-2
Azinphos-methyl	86-50-0
Bromopropylate	18181-80-1
Chlordane (cis and trans)	57-74-9
Chlorfenvinphos	470-90-6
Chlorpyrifos	2921-88-2
Chlorpyrifos-methyl	5598-13-0
Cypermethrin	52315-07-8
DDT	50-29-3
Deltamethrin	52918-63-5
Diazinon	333-41-5
Dichlorvos	62-73-7
Dieldrin	50-57-1
Dithiocarbamates	142-84-7
Endosulfan	115-29-7
Endrin	72-20-8
Ethion	563-12-2
Fenitrothion	122-14-5
Fenvalerate	51630-58-1
Fonofos	944-22-9
Heptachlor	76-44-8
Hexachlorobenzene	118-74-1
Hexachlorocyclohexane	608-73-1
Lindane	58-89-9
Malathion	121-75-5
Methidathion	950-37-8
Parathion	56-38-2
Parathion-methyl	298-00-0
Permethrin	52645-53-1
Phosalone	2310-17-0
Piperonyl butoxide	51-03-6
Pirimiphos-methyl	29232-93-7
Pyrethrins	8003-34-7
Quintozene (sum of 3 items)	82-68-8

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Protein SDS Gel Electrophoresis

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Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10174

Lot #: N230927E

Sponsor: *Active Concepts, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principal Investigator: *Hannah Duckett*

Test Performed:
Protein SDS Gel Electrophoresis

Introduction

Molecular weight is a critical component of topical cosmetic products given compounds with larger molecular weights do not penetrate the skin as deep as smaller molecular weight compounds. Molecular weights can also be utilized to determine the impact of cosmetic products on specific molecules of interest. Specifically, the relative amount of a molecule within a given range can be assessed, providing insight to an increase or decrease in particular molecules that impact the overall function of skin differently.

Accordingly, Protein SDS Gel Electrophoresis was performed to determine the *in vitro* effect of **FortiCulture Coconut** on proteins within the 4 – 10 kDa range.

Assay Principle

In vertical gel electrophoresis the wells are loaded in a vertical position and as the current runs through the gel the bands, visible due to the loading dye, move through the gel top to bottom. As the current increases, the speed at which the bands move through the gel increases. This can lead to band warping and lack of distinct band formation. Due to this, moderate currents with longer run times are generally chosen for better downstream results. When samples are run with known standard molecular weight ladders, gel electrophoresis becomes a beneficial tool for determining molecular weight of a sample. Running protein samples alongside one another also demonstrates a visual comparison of band migration and intensity sample to sample. Protein gel electrophoresis provides both quantitative as well as qualitative data for protein samples of interest.

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Materials

A. Run Conditions:	125 Volts for 55 Minutes
B. Incubation Conditions:	37°C at 5% CO_2 and 95% Relative Humidity (RH)
C. Equipment:	Forma Humidified Incubator; ESCO Biosafety Cabinet; Light Microscope; Mini Gel Tank (Invitrogen); Power Supply (Hoefer); Pipettes; Canon Powershot SX160IS Digital camera
D. Cell Line:	Normal Human Epidermal Keratinocytes (ATCC; PCS-200-011)*
E. Media/Buffers:	Keratinocyte Basal Medium (ATCC; PCS-200-030)*; Keratinocyte Growth Kit (ATCC; PCS-200-040)*; Phosphate Buffered Saline (PBS)
F. Culture Plate:	6-Well Tissue Culture Treated Plate
G. Gel:	12-well 10% Polyacrylamide
H. Reagents:	SureCast Acrylamide 40%*; SureCast Resolving Buffer; Distilled Water; 10% SureCast APS*; SureCast TEMED*; Simply Blue SafeStain*; NuPAGE SDS Running Buffer (20X)*; Native Tris-Glycine Sample Buffer (2X)*
I. Other:	Spectra Multicolor Low Range Protein Ladder 42-1.7 kDa (Thermo Scientific)*; Sterile disposable pipette tips; Microcentrifuge tubes
J. Software:	ImageJ (National Institutes of Health); Excel Analysis ToolPak (Microsoft)

**Or suitable alternatives, subject to change without notice based off vendor availability*

Methods

Human epidermal keratinocytes from the same lot were seeded into a 6-well tissue culture microplate and grown to 80%-90% confluency in Complete Media at 37°C. Keratinocytes were incubated with Complete Media in addition to 0.01% and 0.04% concentrations of **FortiCulture Coconut** in Complete Media. Following a 72-hour incubation, the media in all wells was removed and cells were washed once with PBS (Phosphate Buffered Saline). 1 mL of cold PBS was added to each well and cells were scraped off the well bottom. The cell suspensions were collected and subjected to two freeze thaw cycles, followed by a centrifuge spin at 5,000 xg for 5 minutes to lyse the cells. The supernatants were collected after the centrifuge spin.

The SureCast plate and gel casting system was used to prepare the polyacrylamide gels. The gels were prepared using the specifications listed in the SureCast system guidelines specific to the particular grade gel chosen for the sample type. A 12-well comb was used to create wells for up to 12 samples to be loaded. Once the gels had hardened, the comb was removed, and the gels were rinsed twice with 1X NuPAGE SDS Running Buffer. The casting plates were locked into the mini gel tank and the remaining running buffer was used to fill the tank. The supernatants were diluted in a 1:8 sample: loading buffer ratio and 15 µL of each sample mixture were added to the designated wells. 15 µL of the Spectra Multicolor Low Range Protein Ladder was added to the designated well.

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The gel was run for 55 minutes at 125V, removed from the glass casting plates, and rinsed with distilled water three times for five minutes each rinse with slight agitation. The gel was then fully submerged in the Simply Blue SafeStain for 2.5 hours with intermittent agitation and then rinsed in distilled water for up to one hour. Digital images of the gel were taken after the water rinse.

ImageJ histogram analysis was utilized to quantify differences in protein content via differences in color intensity. A region of interest was designated for each sample in the molecular weight range of the proteins of interest and a histogram was generated for each area (all regions of interest were the same size between samples). The percent increase in molecular weight was calculated relative to the Complete Media control. Three separate experiments were performed and average values were recorded. Data was analyzed using a one-way ANOVA with statistical significance accepted at $p \leq 0.05$.

Results

The data obtained from this study met criteria for a valid assay; the standard ladders performed as anticipated.

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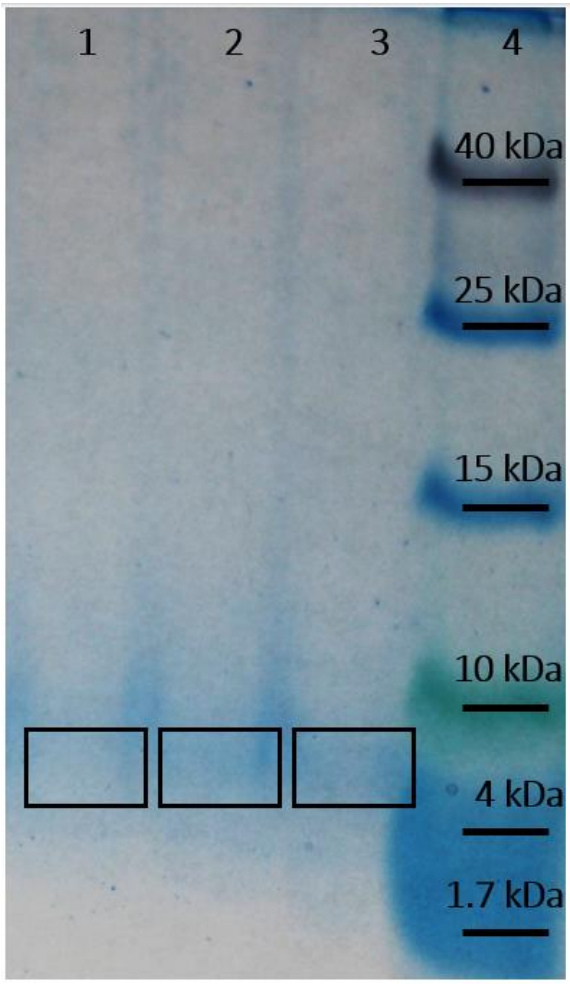

1. Complete Media
2. 0.01% FortiCulture Coconut
3. 0.04% FortiCulture Coconut
4. Low Molecular Weight Range Protein Ladder

Figure 1: Final stained gel comparing **FortiCulture Coconut** to the low molecular weight range protein ladder

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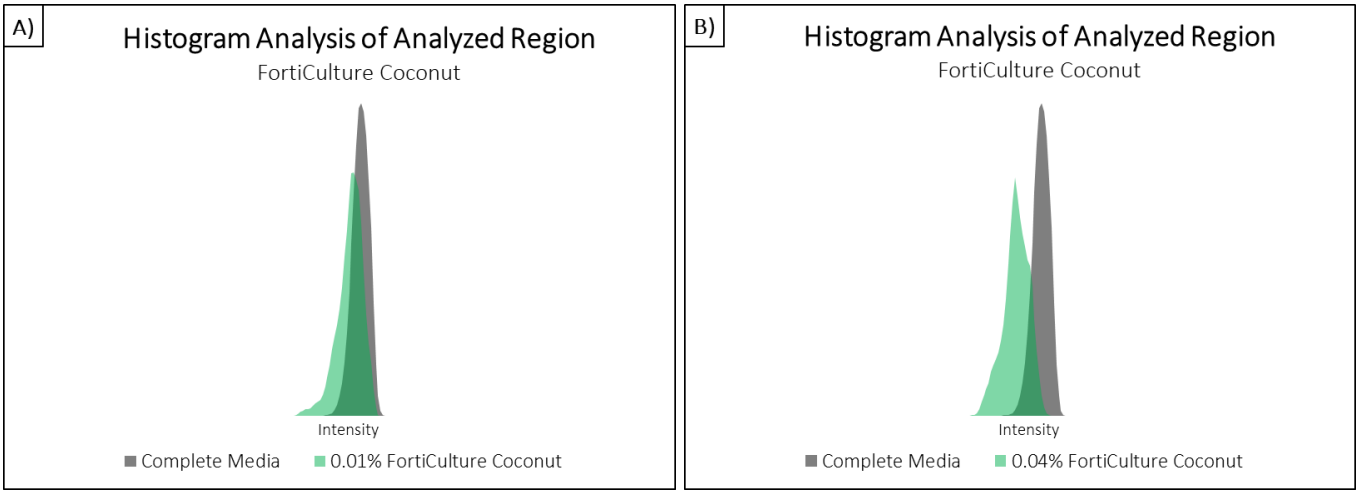

Figure 2: Histograms of the analyzed regions as shown in Figure 1. A) Histograms of the analyzed regions for Complete Media and 0.01% **FortiCulture Coconut**. B) Histograms of the analyzed regions for Complete Media and 0.04% **FortiCulture Coconut**.

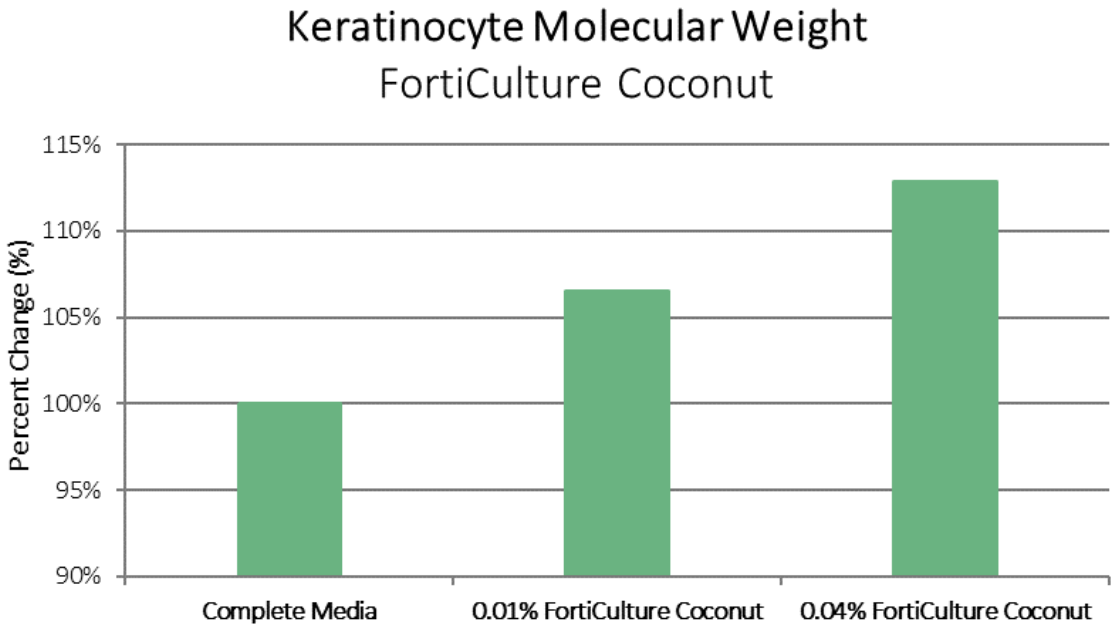

Figure 3. Percent Change in Keratinocyte Molecular Weight Relative to Complete Media

Table 1. P-values of One-way ANOVA Statistical Analysis

	Complete Media vs 0.01% **FortiCulture Coconut**	Complete Media vs 0.04% **FortiCulture Coconut**
P-value	0.041	0.033

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Discussion

The migration rate of the bands directly correlates with the molecular weight of the sample material. The larger the molecular weight of a sample, the heavier the band is and the shorter distance it will travel down the gel. In contrast, the smaller the molecular weight of the sample, the lighter the band is and the further distance it will travel down the gel. A high amount of protein content will exhibit a darker blue and will read as lower pixel intensity.

As shown in Figure 1, the **FortiCulture Coconut** samples and the complete media control regions of interest all migrated to between 4 and 10 kDa. As displayed in Figure 2, 0.01% and 0.04% **FortiCulture Coconut** both exhibit lower luminosity intensity readings than the complete media control which is indicative of higher protein content. With this data we can conclude that **FortiCulture Coconut** was able to increase keratinocyte molecular weight within the region of interest by 7% and 13% at 0.01% and 0.04% use levels, respectively, without increasing protein size (Figures 2, 3). Please note the *in vitro* concentration of 0.01% translates to a 1.0% concentration *in vivo* use-level.

Inhibition Activity Data

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Product Name: FortiCulture Coconut
Code Number: M11001
Lot Number: N220927E
Test Request Number: 9988
CAS #'s: 7732-18-5 & 8001-31-8 & 8013-01-2
EINECS #'s: 231-791-2 & 232-282-8 & 232-387-9
INCI Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate

Organism (ATCC #)	Minimum Inhibitory Concentration (%)
E.coli #8739	0.50
S. aureus #6538	1.00
P. aeruginosa #9027	0.50
C. albicans #10231	1.00
A. brasiliensis #16404	1.00

QA Signature Monica Beltran

Date February 27, 2023

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Antimicrobial Efficacy Test
PCPC Section 20
Method 3

Determination of Preservation Adequacy of Water- Miscible Personal Care Products

Product

M1001-FortiCulture Coconut

Test Request Form

9682

Purpose

This study was initiated to determine the efficacy of a cosmetic ingredient with antimicrobial properties in a Generic Cream Base formula against bioburden as a function of time.

Study Dates

The study was started on November 16th, 2022 and was completed on January 20th, 2023.

Test Organisms

Gram-positive Bacteria	*Staphylococcus aureus* ATCC# 6538
Gram-negative Bacteria	*Escherichia coli* ATCC# 8739
	Pseudomonas aeruginosa ATCC# 9027
Mold and Yeast	*Aspergillus brasiliensis* ATCC# 16404
	Candida albicans ATCC# 10231

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Neutralization:

Verification of neutralization of the antimicrobial properties of the product was demonstrated prior to performing the test for microbial content by inoculating the product dilution with a low level of challenge microorganisms (100 CFU) and verifying recovery of this viable inoculum using Dey/Engley (D/E) broth. This provides evidence that the antimicrobial has been neutralized and there are no false positive results during the Challenge Test.

Test Method

Fifty grams of Generic Cream Base formula pH 7 (Table 2) with 1.0% FortiCulture Coconut was weighed into five individual containers. Each container was inoculated with one of the five test organisms. The inoculum concentration for each organism was standardized using the 0.5 McFarland turbidity standard and further diluted to yield approximately 10^6 to 10^8 microorganisms/ml. The amount of each inoculum added to each sample was no more than 1% of the product weight, as to not alter the product composition.

The inoculated samples were evaluated 0, 7, 14, 21, and 28 days after the initial inoculation to determine quantitatively the number of viable microorganisms remaining. On the 28th day of testing the samples were re-inoculated and evaluated 7, 14, 21, and 28 days after the second exposure to determine the number of viable microorganisms. Table 1 represents the percent reduction of viable organisms after being introduced into the test formulation.

Results & Discussion

The results obtained from the Neutralization Test of each product using Dey/Engley (D/E) broth, indicate that the neutralization steps conducted prior to performing the Challenge Test are indeed effective for avoiding false positive Challenge Test results.

The results of this Challenge Test (Table 1) demonstrate the effectiveness of the preservation system used in Generic Cream Base formula pH 7 with 1.0% FortiCulture Coconut. The recommendations stated in Section 13, Determination of Preservative Adequacy in Cosmetic Formulations, in the PCPC Microbiology Guidelines are as follows:

Bacteria – There should be at least a 99.9% (3 log) reduction of vegetative bacteria within 7 days following each challenge and no increase for the duration of the test period.

Yeasts and Molds – There should be at least a 90% (1 log) reduction of yeasts and molds within 7 days following each challenge and no increase for the duration of the test period.

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Percentage Reduction of Viable Organisms					
Inoculum (initial) CFU/ml	*E. coli*	*P. aeruginosa*	*S. aureus*	*A. brasiliensis*	*C. albicans*
	6.0×10^6	3.0×10^6	4.0×10^6	1.1×10^6	4.2×10^5
Day 0*	99.826%	99.976%	99.950%	99.993%	99.914%
Day 7	99.997%	99.984%	99.995%	99.999%	99.999%
Day 14	>99.999%	>99.999%	>99.999%	99.999%	99.999%
Day 21	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%
Day 28	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%
Inoculum (re-inoculated) CFU/ml	3.0×10^6	3.1×10^6	6.0×10^6	1.2×10^6	2.0×10^5
Day 7	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%
Day 14	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%
Day 21	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%
Day 28	>99.999%	>99.999%	>99.999%	>99.999%	>99.999%

Table 1. Challenge Test results for Generic Cream Base formula pH 7 with 1.0% FortiCulture Coconut
Inoculated on Day 0 and re-inoculated on Day 28. Results show % reduction in viable organisms.

* The days listed in the first column refer to the inoculum/plating day. Bacteria results are read 2 days after plating day, and mold and yeast results are read 5 days after plating day.

Conclusions:

Under the conditions of this test, Gram positive and Gram negative bacteria as well as the yeast and mold were reduced by greater than 99.9% within 7 days of each challenge. By the end of each 28-day test period Gram positive and Gram negative bacteria as well as the yeast and mold were reduced by 99.999% or greater.

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Phase	Ingredient	Supplier	%
I	Water	-	85.2
	Carbopol Ultrez 10	Lubrizol	0.1
	Glycerin	PT. Musim Mas	3.0
	Tealan	RITA	0.9
II	Cetyl Alcohol	RITA	2.0
	Stearic Acid	Acme Hardesty Oleochemicals	0.8
	Glyceryl Stearate	Protameen Chem.	1.5
	Isopropyl Myristate	Alzo	1.5
	Mineral Oil	RITA	5.0

Table 2. Generic Cream Base (O/W) formula

Manufacturing Process:

1. Phase I:
Charge water into the main beaker and begin propeller mixing. A vortex should form. Begin heating to 75^{0}C. Slowly sift in Carbopol while mixing. Add the rest of the ingredients.

2. Phase II:
In a separate beaker, combine ingredients and heat to 75^{0}C while mixing. Mix until homogenous. Then add to the main beaker with high-speed mixing. Maintain temperature at 75^{0}C and mix for 30 minutes. Begin force cooling to 25^{0}C.

3. Check the pH and adjust as necessary with Citric Acid (50%) or NaOH (25%).

Specifications:

Appearance: White to Off-White Emulsion
pH: 6.5 – 8.0
*If a different pH is desired, adjust using Citric Acid (50%) or NaOH (25%). Formula is stable in the 3.0 – 7.0 pH range.

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Antimicrobial Efficacy (Challenge) Testing

The intent of performing an Antimicrobial Efficacy or Challenge test is to evaluate whether an antimicrobial agent or preservation system in a given cosmetic formulation has the ability to prevent the growth of test microorganisms. The test methodology employed by Active Micro Technologies (AMT) is based on the methods published in the CTFA Microbiology Guidelines. AMT's goal is to assist our customers by providing a screening test of a product formulation that is approaching finalization. It is expected that the formulation(s) submitted for Challenge testing contain AMT antimicrobials and have already passed the customer's internal stability tests. It is also anticipated that formal challenge testing of the final formulation will subsequently be performed by the customer at an outside lab of their choosing.

The information contained in this report is provided by Active Micro Technologies after the exercise of all reasonable care and skill in its compilation, preparation, and issue. It is provided without liability regarding its subsequent application and use. This type of screening test will be conducted only for validation of the efficacy of the antimicrobial agent or preservative system in the specific formulation tested. It does not address the suitability of the overall formula, nor does it address the regulatory status of any component therein. This testing does not account for the possibility of environmental microorganisms and cannot be relied upon as sufficient to justify commercialization of the product tested. By submitting samples for testing, the customer acknowledges that they will not hold Active Micro Technologies responsible for products launched based solely on the support of these studies.

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Reactive Oxygen Species Scavenging Assay

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 9906

Lot #: N220927E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Daniel Shill*

Test Performed:
Reactive Oxygen Species Scavenging Assay

Introduction

Low levels of intracellular oxidative stress are produced during normal physiological functions. However, UV irradiation, pollutants, foreign substances, and aging elicit unrestricted increases in reactive oxygen species (ROS). These deregulated augmentations in oxidative stress lead to an acceleration of DNA mutation, cellular senescence, advanced glycation end products, protein oxidation, and collagen degradation. Moreover, when intrinsic antioxidant capacities are reduced, such as during aging, an imbalance between pro- and anti-oxidant systems further accentuates these hallmarks of cellular aging.

Accordingly, a ROS Scavenging Assay was conducted to assess the *in vitro* effect of **FortiCulture Coconut** to scavenge unnecessary oxidative stress in dermal fibroblasts. Attenuating excessive ROS preserves cellular homeostasis and blunts intrinsic and extrinsic age-related declines in skin cell function.

Assay Principle

Two cell-permeant dyes, CellROX™ Orange Reagent and Hoechst, were utilized in conjunction to provide a specific and quantitative method for determining ROS levels. CellROX™ Orange Reagent fluoresces brightly when bound to ROS indicating oxidative stress, and Hoechst fluoresces when bound to nuclear DNA to indicate cellular nuclei. By displaying the relative fluorescent units (RFU) from the CellROX™ Orange Reagent (ROS Signal) as a function of Hoechst (Nuclear Signal), ROS can be quantified and normalized at the cellular level. To elicit supraphysiological mitochondrial- and non-mitochondrial-derived levels of oxidative stress, the cells were exposed to Antimycin A, a complex III inhibitor of the mitochondrial electron transport chain.

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Materials

A. Kit:	CellROX™ Orange Reagent (ThermoFisher Scientific, C10443)
B. Incubation Conditions:	37°C at 5% CO_2 and 95% Relative Humidity (RH)
C. Equipment:	Forma humidified incubator; ESCO biosafety laminar flow hood; Light microscope; Synergy HT Microplate Reader; Pipettes
D. Cell Line:	Normal Human Neo-Natal Dermal Primary Fibroblasts (ATCC, PCS-201-010)
E. Media/Buffers:	Fibroblast Basal Medium (PCS-201-030); Fibroblast Growth Kit (ATCC, PCS-201-041); Ethanol; Phosphate Buffered Saline (PBS)
F. Reagents:	Hoechst 33342 (ThermoFisher Scientific, 62249); Antimycin A (Sigma Aldrich, A8674)
G. Culture Plate:	96 Well Black Side/Clear Bottom Tissue Culture Treated Microplates
H. Software:	Excel Analysis ToolPak (Microsoft)
I. Other:	Sterile disposable pipette tips

Methods

Human dermal fibroblasts were seeded into a 96-well tissue culture microplate and grown to 80%-90% confluency in complete media (CM). 0.01%, 0.1% and 1.0% concentrations of **FortiCulture Coconut** in CM were added to cells and placed at 37°C. Control wells were incubated with CM only. Following an 18-hour incubation, the media in all wells was removed and cells were washed once with PBS. Hoechst and CellROX™ Orange were diluted in CM, and added to all wells at final concentrations of 20 µM and 5 µM, respectively. Following a 30-minute incubation at 37°C, the Hoechst and CellROX™ Orange solution was removed and cells were washed once with PBS. Next, 200 pM of Antimycin A (AntA), initially dissolved in ethanol and further diluted in CM, was added to all wells, except control wells that received CM. Following another 30-minute incubation at 37°C, the AntA and CM was removed, CM was added to all wells, and fluorescence measurements were taken with the following wavelengths (excitation / emission): Hoechst (361 nm / 486 nm) and CellROX™ Orange (545 nm / 565 nm).

Three separate experiments were performed with conditions in duplicate and average values were recorded. Data was analyzed using a one-way ANOVA with statistical significance accepted at $p \leq 0.05$. To account for differences in cell counts, ROS levels are expressed as the ROS Signal (CellROX™ Orange) divided by the Nuclear Signal (Hoechst), as calculated by the following equation:

$$ROS\ Levels = \frac{ROS\ Signal}{Nuclear\ Signal}$$

Percent change is expressed relative to AntA and calculated by the following equation:

$$Percent\ Change\ (\%) = \frac{RFU_{Sample} - RFU_{AntA}}{RFU_{AntA}} \times 100$$

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Results

The data obtained from this study met criteria for a valid assay and the positive control performed as anticipated. Compared to untreated fibroblasts, AntA (200 pM) increased ROS levels. Fibroblasts treated with AntA and **FortiCulture Coconut** at 0.01%, 0.1%, and 1.0% exhibited a reduction in oxidative stress levels compared to fibroblasts exposed to AntA.

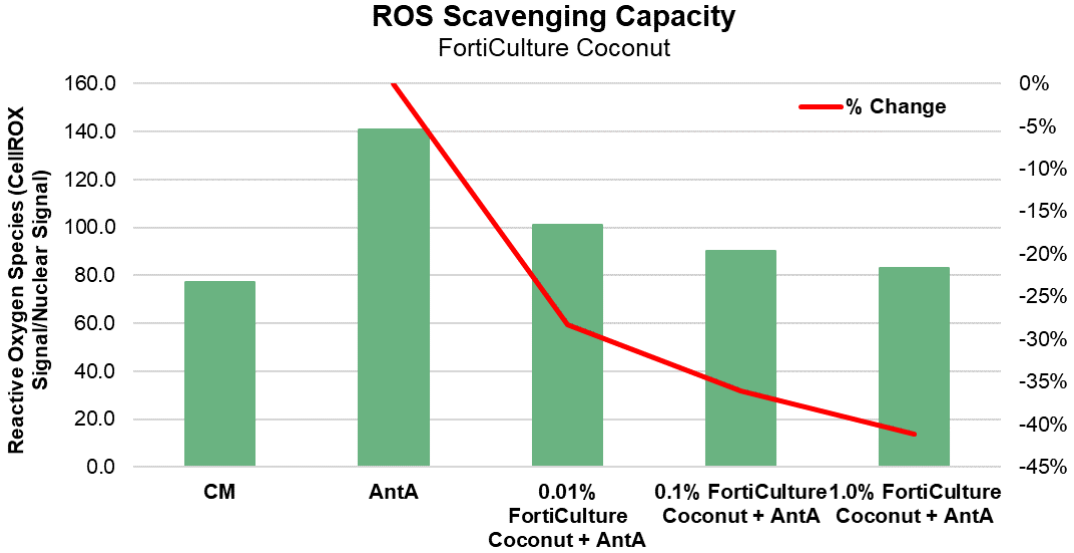

Figure 1: The effect of **FortiCulture Coconut** on ROS scavenging.

Table 1: P-values from one-way ANOVA Statistical Analysis Compared to AntA Treated Fibroblasts

	Complete Media	0.01% **FortiCulture Coconut**	0.1% **FortiCulture Coconut**	1.0% **FortiCulture Coconut**
P-value	< 0.001	0.007	0.002	< 0.001

Discussion

As shown in Figure 1, fibroblasts incubated with AntA, a known inducer of oxidative stress, elicited an 82% increase in ROS levels, compared to untreated fibroblasts. These data demonstrate the supraphysiologic level of ROS induced by AntA and the magnitude of ROS in fibroblasts is dynamic.

Conversely, fibroblasts treated with AntA and **FortiCulture Coconut** at 0.01%, 0.1%, and 1.0% demonstrated 28%, 36%, and 41% reductions in ROS levels compared to fibroblasts treated with AntA, respectively. These data demonstrate **FortiCulture Coconut** attenuates excessive oxidative stress.

Collectively, intrinsic and extrinsic factors perturb skin homeostasis by stimulating abundant levels of ROS that amplify DNA mutation, cellular senescence, advanced glycation end products, protein oxidation, and collagen degradation. These data indicate **FortiCulture Coconut** scavenges unnecessary ROS, which may help to attenuate characteristics of cellular aging.

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SA-β-gal Activity Cellular Aging Model Fibroblasts

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10172

Lot #: N220927E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Daniel Shill*

Test Performed:
Senescence-Associated β-galactosidase (SA-β-gal) Activity
Cellular Aging Model: Fibroblasts

Introduction

Cellular senescence is a state of permanent cell cycle arrest that accompanies aging and contributes to a decline in normal skin function and physiology. Contrary to apoptosis, defined as programmed cell death resulting in the clearance of damaged cells, senescent cells are stable, viable, and communicate with neighboring cells. Senescence is characterized by changes in cellular morphology, metabolism, signaling pathways, and biochemical profiles that manifest as increased collagen breakdown, wrinkles, and thin skin. SA-β-gal is the gold standard biomarker to identify senescence *in vitro* as the enzyme β-galactosidase explicitly accumulates in the lysosomes of senescent cells.

Accordingly, a cellular aging model was developed to assess the *in vitro* effect of **FortiCulture Coconut** to reduce SA-β-gal activity in “aged” fibroblasts. Attenuating cellular senescence could blunt or prevent the age-related decline in skin function and physiology.

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Assay Principle

A model of cellular aging was developed by utilizing fibroblasts at different passage numbers (P), which represent the number of times cells have been harvested and reseeded into subsequent cell culture vessels (i.e. the higher the passage number, the more times those cells have been harvested and reseeded). Fibroblasts at low passages (≤ P5) demonstrate morphological, metabolic, and senescent biomarker profiles matching that of young skin *in vivo*. Conversely, fibroblasts at high passages (≥ P8) exhibit the cellular characteristics of older skin *in vivo*.

Hoechst and SPiDER-ßGal dyes were utilized in conjunction to provide a specific and quantitative method for determining cellular senescence. Hoechst, a cell-permeant dye, fluoresces brightly when bound to nuclear DNA to indicate cellular nuclei, whereas SPiDER-ßGal fluoresces when bound to β-galactosidase indicating cellular senescence. SA-β-gal activity is normalized by displaying the relative fluorescent units (RFU) from the SPiDER-ßGal dye (β-galactosidase Signal) as a function of the Hoechst dye (Nuclear Signal) RFU and set relative to “young” fibroblasts to underscore the effect of cellular aging on senescence.

Materials

A. Kit:	Cellular Senescence Plate Assay Kit – SPiDER-ßGal (Dojindo; SG05)*
B. Incubation Conditions:	37°C at 5% CO_2 and 95% Relative Humidity (RH)
C. Equipment:	Forma Humidified Incubator; ESCO Biosafety Cabinet; Light Microscope; Synergy HT Microplate Reader; Pipettes
D. Cell Line:	Normal Human Dermal Fibroblasts (ATCC; PCS-201-012)*
E. Media/Buffers:	Fibroblast Basal Medium (ATCC; PCS-201-030)*; Fibroblast Growth Kit (ATCC; PCS-201-041)*; Phosphate Buffered Saline (PBS)
F. Reagents:	Hoechst 33342 (ThermoFisher Scientific; 62249)*; Dimethyl Sulfoxide (DMSO)
G. Culture Plate:	96-Well Black Side/Clear Bottom Tissue Culture Treated Microplates
H. Software:	Excel Analysis ToolPak (Microsoft)
I. Other:	Sterile disposable pipette tips

*Or suitable alternatives, subject to change without notice based off vendor availability

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Methods

"Young" and "aged" human dermal fibroblasts from the same lot were seeded into a 96-well tissue culture microplate and grown to 80%-90% confluency in complete media (CM) at 37°C. "Young" fibroblasts were utilized in this assay at passage P4, while "aged" fibroblasts were utilized at passage P8. "Young" and "aged" fibroblasts incubated with CM only are utilized as controls, whereas 0.01% and 0.1% concentrations of **FortiCulture Coconut** in CM were added to "aged" fibroblasts only. Following a 24-hour incubation, the media in all wells was removed and cells were washed once with PBS. The nuclear dye Hoechst was diluted in CM, added to all wells at final concentration of 10 µM, and incubated at 37°C.

Following a 30-minute incubation, the Hoechst dye was removed, all cells were washed once with PBS, CM was added to all wells, and fluorescence measurements were taken to determine the Nuclear Signal (excitation: 361 nm / emission: 486 nm). Subsequently, the CM was removed, all cells were washed once with PBS, and Lysis Buffer was added to each well. After a 10-minute incubation at room temperature, the SPiDER-ßGal dye was added to each well and incubated for 30 minutes at 37°C. The Stop Solution was then added to each well and fluorescence measurements were taken to determine the β-galactosidase Signal (excitation: 535 nm / emission: 580 nm).

Three separate experiments were performed with conditions in duplicate and average values were recorded. Data was analyzed using a one-way ANOVA with statistical significance accepted at $p \leq 0.05$. To account for differences in cell counts, normalized SA-β-gal activity is calculated as the β-galactosidase Signal (SPiDER-ßGal dye) divided by the Nuclear Signal (Hoechst dye), as shown by the following equation:

$$Normalized\ SA\text{-}\beta\text{-}gal\ Activity = \frac{\beta\text{-}galactosidase\ Signal}{Nuclear\ Signal}$$

SA-β-gal activity is displayed as relative to the normalized SA-β-gal activity in "young" fibroblasts by the following equation:

$$SA\text{-}\beta\text{-}gal\ Activity = \frac{Normalized\ SA\text{-}\beta\text{-}gal\ Activity_{Sample}}{Normalized\ SA\text{-}\beta\text{-}gal\ Activity_{Young\ Fibroblasts}}$$

Percent change is calculated from the SA-β-gal activity in "aged" fibroblasts and calculated by the following equation:

$$Percent\ Change\ (\%) = \frac{SA\text{-}\beta\text{-}gal\ Activity_{Sample} - SA\text{-}\beta\text{-}gal\ Activity_{Aged\ Fibroblasts}}{SA\text{-}\beta\text{-}gal\ Activity_{Aged\ Fibroblasts}} \times 100$$

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Results

The data obtained from this study met criteria for a valid assay and the controls performed as anticipated. Compared to “aged” fibroblasts, “young” fibroblasts demonstrated lower levels of SA-β-gal activity. “Aged” fibroblasts treated with **FortiCulture Coconut** at 0.01% and 0.1% exhibited a reduction in SA-β-gal activity compared to untreated “aged” fibroblasts.

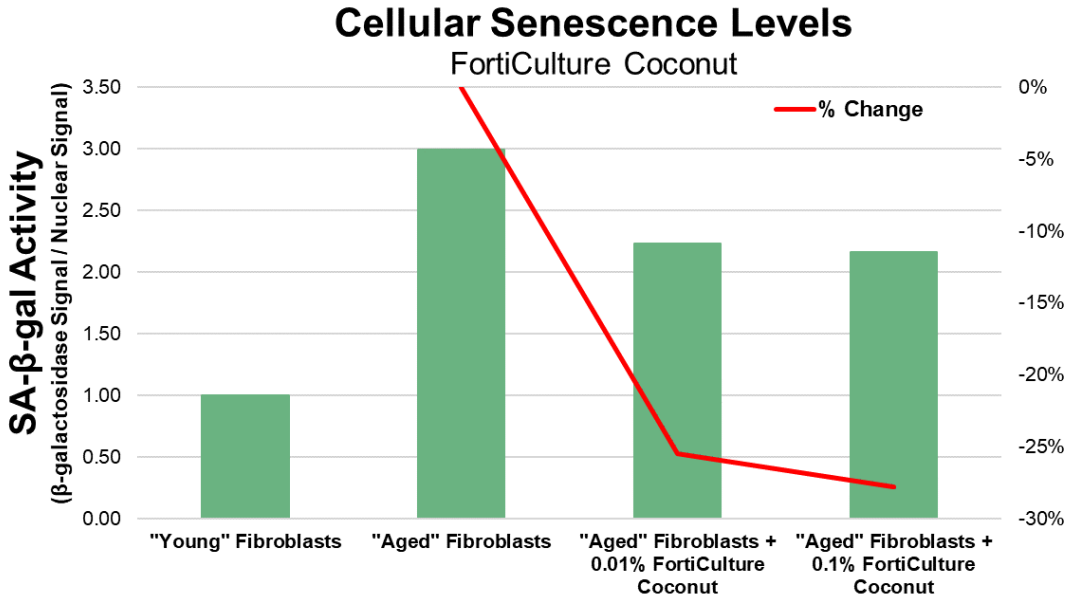

Figure 1: The effect of **FortiCulture Coconut** on cellular senescence levels in “aged” fibroblasts.

Table 1: P-values from one-way ANOVA statistical analysis compared to “aged” fibroblasts.

	“Young” Fibroblasts	“Aged” Fibroblasts + 0.01% **FortiCulture Coconut**	“Aged” Fibroblasts + 0.1% **FortiCulture Coconut**
P-value	< 0.001	0.013	0.025

Discussion

As shown in Figure 1, “aged” fibroblasts demonstrated 199% higher levels of SA-β-gal activity compared to “young” fibroblasts. These data demonstrate “young” fibroblasts exhibit reduced levels of cellular senescence compared to “aged” fibroblasts. However, “aged” fibroblasts treated with **FortiCulture Coconut** at 0.01% and 0.1% elicited 25% and 28% reductions in SA-β-gal activity compared to untreated “aged” fibroblasts, respectively. These data demonstrate **FortiCulture Coconut** attenuates cellular senescence in “aged” fibroblasts.

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Senescence is a hallmark characteristic of cellular aging that contributes to the age-associated modifications of skin function and composition *in vivo*, such as increased dermal thinning, loss of elasticity, and the development of wrinkles. Specifically, cellular senescence is associated with increased inflammation, oxidative stress, and impaired collagen homeostasis. Collectively, these data indicate that **FortiCulture Coconut** reduces cellular senescence and may attenuate or reverse the alterations in skin structure and physiology that occur during aging.

Sirius Red/Fast Green Collagen Analysis

107 Technology Drive • Lincolnton, NC 28092
(704) 276-7100 • Fax (704) 276-7101

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10173

Lot #: N220927E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principal Investigator: *Hannah Stade*

Test Performed:
Sirius Red/Fast Green Collagen Assay

Introduction

Collagen is the main protein of connective tissues, such as skin, bone, tendon and ligament, and the most abundant protein in mammals. Specifically, it accounts for nearly 25% to 35% of the total human protein content. Collagen is a long, fibrous protein that forms bundles called fibers giving structure and support to cells and tissues. Collagen has great tensile strength and is responsible for skin's elasticity, therefore degradation leads to wrinkles that accompany aging.

Accordingly, a Sirius Red/Fast Green Collagen Assay was conducted to assess the *in vitro* effect of **FortiCulture Coconut** to trigger collagen synthesis in dermal fibroblasts.

Assay Principle

Sirius Red is a unique dye that binds specifically to the helical structure of types I through V collagen, while Fast Green binds to non-collagenous proteins. These two dyes work in conjunction to provide a semi-quantitative method of determining amounts of collagen and non-collagenous proteins in a sample. After staining samples, the dyes are easily extracted and have optical density (OD) absorptions at 540 nm (Sirius Red) and 605 nm (Fast Green). Collagen concentrations are calculated through equations with OD values.

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Materials

A. Kit:	Sirius Red/Fast Green Collagen Kit (Chondrex; 9046)*
B. Incubation Conditions:	37°C, 5% CO_2, and 95% relative humidity
C. Equipment:	Forma humidified incubator; ESCO biosafety laminar flow hood; Synergy HT Microplate reader; Pipettes; Light microscope
D. Cell Line:	Normal Human Dermal Fibroblasts (ATCC; PCS-201-012)*
E. Media/Buffers:	Fibroblast Basal Medium (PCS-201-030)*; Fibroblast Growth Kit (PCS-201-041)*; Phosphate Buffered Saline (PBS)
F. Reagents:	Sirius Red/Fast Green dye solution (Chondrex; 9046)*; Extraction solution (Chondrex; 9046)*; Ascorbic Acid-2-Glucose (AA2G) (100 µM or 34 µg/mL); Insulin Growth Factor-1 (IGF-1) (6.5 nM or 50 ng/mL); Glacial Acetic Acid; Ethanol
G. Culture Plate:	Flat Bottom 24-Well Tissue Culture Treated Plates
H. Other:	Sterile disposable pipette tips

**Or suitable alternatives, subject to change without notice based off vendor availability*

Methods

Human dermal fibroblasts were seeded into 24-well tissue culture plates and allowed to grow to confluency in Complete Media. 0.01%, 0.02%, and 0.04% concentrations of **FortiCulture Coconut** were diluted in serum-free Complete Media and incubated with fibroblasts for 24 hours. Ascorbic Acid-2-Glucose (AA2G) (100 µM or 34 µg/mL) and Insulin Growth Factor-1 (IGF-1) (6.5 nM or 50 ng/mL) were diluted in serum-free Complete Media and utilized as positive controls.

Media was removed from wells containing adherent fibroblasts and the cells were washed with PBS. 500 µL of a cooled 95% ethanol/5% glacial acetic acid solution was added to the wells and incubated for 10 minutes at room temperature, followed by a PBS wash. 200 µL of the Sirius Red/Fast Green dye solution was added to the fixed cell layer and incubated at room temperature for 30 minutes. The dye solution was removed from the cell layer and washed with PBS. 1 mL of extraction solution was added for color extraction and optical density (OD) was read at 540 nm and 605 nm on a Synergy HT Microplate Reader.

Three separate experiments were performed with conditions in duplicate and average values were recorded. Data was analyzed using a one-way ANOVA with statistical significance accepted at $p \leq 0.05$. The collagen and non-collagen protein concentrations of fibroblasts were determined by the following calculations and expressed in µg:

$$Collagen\ (\mu g) = \frac{OD_{540} - (OD_{605} \times 0.291)}{0.0378}$$

$$Non\ Collagen\ Protein\ (\mu g) = \frac{OD_{605}}{0.00204}$$

Percent change is expressed relative to untreated fibroblasts (Complete Media) and calculated by the following equation:

$$Percent\ Change\ (\%) = \frac{Collagen_{Sample} - Collagen_{Complete\ Media}}{Collagen_{Complete\ Media}} \times 100$$

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Results

The data obtained from this study met criteria for a valid assay and the positive controls performed as anticipated. Compared to untreated fibroblasts, AA2G (100 µM or 34 µg/mL) and IGF-1 (6.5 nM or 50 ng/mL) increased collagen concentrations. Fibroblasts treated with **FortiCulture Coconut** at 0.01%, 0.02%, and 0.04% demonstrated increased collagen synthesis compared to untreated fibroblasts.

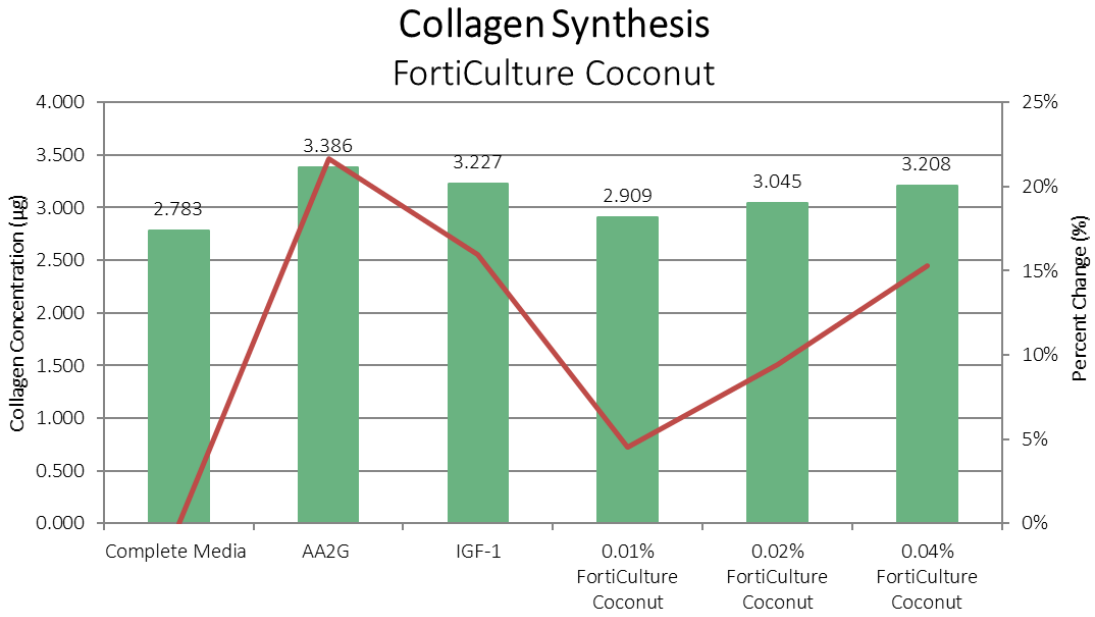

Figure 1. The effect of AA2G (100 µM or 34 µg/mL), IGF-1 (6.5 nM or 50 ng/mL), and **FortiCulture Coconut** on collagen concentrations in dermal fibroblasts.

Table 1. Results from one-way ANOVA Statistical Analysis Compared to Complete Media

	0.01% **FortiCulture Coconut**	0.02% **FortiCulture Coconut**	0.04% **FortiCulture Coconut**
P-value	0.041	0.030	0.026

Discussion

As shown in Figure 1, fibroblasts incubated with AA2G and IGF-1, both known to stimulate collagen synthesis, exhibited increases in collagen synthesis of 22% and 16% compared to untreated fibroblasts, respectively. These data demonstrate collagen in fibroblasts is dynamic and can be manipulated with exogenous compounds. Similarly, fibroblasts treated with **FortiCulture Coconut** at 0.01%, 0.02%, and 0.04% demonstrated potent increases in collagen synthesis of 5%, 9%, and 15%, respectively, compared to untreated fibroblasts. Overall, these data demonstrate **FortiCulture Coconut** activates collagen synthesis in dermal fibroblasts.

Collectively, increases in collagen production may lead to improvements in dermal-epidermal junction integrity as well as an improved scaffolding matrix. These data indicate **FortiCulture Coconut** stimulates collagen synthesis which may assist cosmetic applications designed to enhance collagen deposition and provide a younger and healthier dermal complexion.

Bacterial Reverse Mutation Test

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(704) 276-7100 • Fax (704) 276-7101

Test Article: FortiCulture Coconut
Code Number: M11001
CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Sponsor:
Active Micro Technologies, LLC
107 Technology Drive
Lincolnton, NC 28092

Study Director: *Maureen Danaher*
Principle Investigator: *Monica Beltran*

Test Performed:
Genotoxicity: Bacterial Reverse Mutation Test

Reference:
OECD471/ISO10993.Part3

Test Request Number: 10926

SUMMARY

A *Salmonella typhimurium/Escherichia coli* reverse mutation standard plate incorporation study described by Ames *et al.* (1975) was conducted to evaluate whether a test article solution **FortiCulture Coconut** would cause mutagenic changes in the average number of reveratants for histidine-dependent *Salmonella typhimurium* strains TA98, TA100, TA1537, TA1535 and tryptophan-dependent *Escherichia coli* strain WP2*uvr*A in the presence and absence of Aroclor-induced rat liver S9. This study was conducted to satisfy, in part, the Genotoxicity requirement of the International Organization for Standardization: Biological Evaluation of Medical Devices, Part 3: Tests for Genotoxicity, Carcinogenicity and Reproductive Toxicity.

The stock test article was tested at eight doses levels along with appropriate vehicle control and positive controls with overnight cultures of tester strains. The test article solution was found to be noninhibitory to growth of tester strain TA98, TA100, TA1537, TA1535 and WP2*uvr*A after Sport Inhibition Screen.

Separate tubes containing 2 ml of molten top agar at 45^0C supplemented with histidine-biotin solution for the *Salmonella typhimurium* strains and supplemented with tryptophan for *Escherichia coli* strain were inoculated with 100 µl of tester strains, 100 µl of vehicle or test article dilution were added and 500 µl aliquot of S9 homogenate, simulating metabolic activation, was added when necessary. After vortexing, the mixture was poured across the Minimal Glucose Agar (GMA) plates. Parallel testing was also conducted with positive control correspond to each strain, replacing the test article aliquot with 50µl aliquot of appropriate positive control. After the overlay had solidified, the plates were inverted and incubated for 48 hours at 37^0C. The mean numbers of revertants of the test plates were compared to the mean number of revertants of the negative control plates for each of the strains tested. The means obtained for the positive controls were used as points of reference.

Under the conditions of this assay, the test article solution was considered to be Non-Mutagenic to *Salmonella typhimurium* tester strains TA98, TA100, TA1537, TA1535 and *Escherichia coli* tester strain WP2*uvr*A. The negative and positive controls performed as anticipated. The results of this study should be evaluated in conjunction with other required tests as listed in ISO 100993, Part 3: Tests for Genotoxicity, Carcinogenicity, and Reproductive Toxicology.

All *Salmonella* tester strain cultures demonstrated the presence of the deep rough mutation (*rfa*) and the deletion in the *uvr*B gene. Cultures of tester strains TA98 and TA100 demonstrated the presence of the Pkm101 plasmid R-factor. All WP2 *uvr*A cultures demonstrated the deletion in the *uvr*A gene. All cultures demonstrated the characteristic mean number of spontaneous revertants in the vehicle controls as follows: TA98, 10-50; TA100, 80-240; TA1535, 5-45; TA1537, 3-21, WP2*uvr*A, 10-60.

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I. Introduction

A. **Purpose**

A *Salmonella typhimurium/Escherichia coli* reverse mutation standard plate incorporation study was conducted to evaluate whether a test article solution would cause mutagenic changes in the average number of revertants for *Salmonella typhimurium* tester strains TA98, TA100, TA1537, TA1535 and *Escherichia coli* WP2*uvr*A in the presence and absences of the S9 metabolic activation. Bacterial reverse mutation tests have been widely used as rapid screening procedures for the determination of mutagenic and potential carcinogenic hazards.

II. Materials

A. **Storage Conditions:** Room temperature (23-25C).
B. **Vehicle:** Sterile DI Water.
C. **Preparation:** Eight different doses level were prepared immediately before use with sterile DI water.
D. **Solubility/Stability:** 100% Soluble and Stable.
E. **Toxicity:** No significant inhibition was observed.

III. Test System

A. Test System

Each *Salmonella typhimurium* and *Escherichia coli* tester strain contains a specific deep rough mutation (*rfa)*, the deletion of *uvr*B gene and the deletion in the *uvr*A gene that increase their ability to detect mutagens, respectively. These genetically altered *Salmonella typhimurium* strains (TA98, TA100, TA1537 and TA1535) and *Escherichia coli* strain (WP2*uvr*A) cannot grow in the absence of histidine and tryptophan, respectively. When placed in a histidine-tryptophan free medium, only those cells which mutate spontaneously back to their wild type states are able to form colonies. The spontaneous mutation rate (or reversion rate) for any one strain is relatively constant, but if a mutagen is added to the test system, the mutation rate is significantly increased.

Tester strain	Mutations/Genotypic Relevance
TA98	hisD3052, Dgal chlD bio *uvr*B *rfa* pKM101
TA100	hisG46, Dgal chlD BIO *uvr*B *rfa* pKM101
TA1537	hisC3076, *rfa*, Dgal chlD bio *uvrB*
TA 1535	hisG46, Dgal chlD bio *uvr*B *rfa*
WP2*uvr*A	trpE, *uvr*A

rfa	=	causes partial loss of the lip polysaccharide wall which increases permeability of the cell to large molecules.
uvrB	=	deficient DNA excision-repair system (i.e., ultraviolet sensitivity)
pKM101	=	plasmid confers ampicillin resistance (R-factor) and enhances sensitivity to mutagens.
*uvr*A	=	All possible transitions and transversions, small deletions.

B. Metabolic Activation

Aroclor induced rat liver (S9) homogenate was used as metabolic activation. The S9 homogenate is prepared from male Sprague Dawley rats. Material is supplied by MOLTOX, Molecular Toxicology, Inc.

C. Preparation of Tester strains

Cultures of *Salmonella typhimurium* TA98, TA100,TA1537, TA1535 and *Escherichia coli* WP2uvrA were inoculated to individual flasks containing Oxoid broth No.2. The inoculated broth cultures were incubated at 37°C in an incubator shaker operating at 140-150 rpm for 12-16 hours.

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D. Negative Control
Sterile DI water (vehicle without test material) was tested with each tester strain to determine the spontaneous reversion rate. Each strain was tested with and without S9 activation. These data represented a base rate to which the number of reveratants colonies that developed in each test plate were compared to determine whether the test material had significant mutagenic properties.

E. Positive Control
A known mutagen for each strain was used as a positive control to demonstrate that tester strains were sensitive to mutation to the wild type state. The positive controls are tested with and without the presence of S9 homogenate.

F. Titer of the Strain Cultures:
Fresh cultures of bacteria were grown up to the late exponential or early stationary phase of growth; to confirm this, serial dilutions from each strain were conducted, indicating that the initial population was in the range of 1 to $2x10^9$/ml.

IV. Method

A. Standard Plate Incorporation Assay:
Separate tubes containing 2 ml of molten top agar supplemented with histidine-biotin solution for the *Salmonella typhimurium* and tryptophan for *Escherichia coli* were inoculated with 100 µl of culture for each strain and 100 µl of testing solution or vehicle without test material. A 500 µl aliquot of S9 homogenate, simulating metabolic activation, was added when necessary. The mixture was poured across Minimal Glucose Agar plates labeled with strain number and S9 activation (+/-). When plating the positive controls, the test article aliquot was replaced by 50µl aliquot of appropriate positive control. The test was conducted per duplicate. The plates were incubated for 37°C for 2 days. Following the incubation period, the revertant colonies on each plate were recorded. The mean number of reveratnts was determined. The mean numbers of revertants of the test plates were compared to the mean number of reverants of the negative control of each strain used.

V. Criteria for a Valid Test

For the test solution to be evaluated as a test failure or “potential mutagen” there must have been a 2-fold or greater increase in the number of mean revertants over the means obtained from the negative control for any or all strains. Each positive control mean must have exhibited at least a 3-fold increase over the respective negative control mean of the *Salmonella* and *Escherichia coli* tester strains used.

All *Salmonella* tester strain cultures must demonstrate the presence of the deep rough mutation (*rfa*) and the deletion in the *uvr*B gene. Cultures of tester strains TA98 and TA100 must demonstrate the presence of the pKM101 plasmid R-factor. All WP2 *uvr*A cultures must demonstrate the deletion in the *uvr*A gene. All cultures must demonstrate the characteristic mean number of spontaneous revertants in the vehicle controls as follows: TA98, 10-50; TA100, 80-240; TA1535, 5-45; TA1537, 3-21, WP2*uvr*A, 10-60. To ensure that appropriate numbers of bacteria are plated, tester strain culture titers must be greater than or equal to $0.3x10^9$ cells/ml. The mean of each positive control must exhibit at least 3.0-fold increase in the number of revertants over the mean value of the respective vehicle control. A minimum of three non-toxicdose levels is required to evaluate assay data. A dose level is considered toxic if one of both of the following criteria are met: (1). A >50% reduction in the mean number of revertants per plate as compared to the mean vehicle control value. This reduction must be accompanied by an abrupt dose-dependent drop in the revertant count. (2). At least a moderate reduction in the background lawn.

VI. Results and Discussion

A. Solubility:
Water was used as a solvent. Solutions from the test article were made from 0.015 to 50mg/ml.

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B. Dose levels tested:
The maximum dose tested was 5000 µg per plate. The dose levels tested were 1.5, 5.0, 15, 50, 150, 500, 1500 and 5000 µg per plate.

C. Titer (Organisms/ml):
5 x 10^8 UFC/ml plate count indicates that the initial population was in the range of 1 to 2 x 10^9 UFC/ml.

D. Standard Plate Incorporation Assay
In no case was there a 2-fold or greater increase in the mean number of revertant testing strains TA98, TA100, TA1537, TA1535 and WP2*uvr*A in the presence of the test solution compared with the mean of vehicle control value. The positive controls mean exhibited at least a 3-fold increase over the respective mean of the *Salmonella typhimurium* and *Escherichia coli* tester strains used. The results are summarized in Appendix 2.

All *Salmonella* tester strain cultures demonstrated the presence of the deep rough mutation (*rfa*) and the deletion in the *uvr*B gene. Cultures of tester strains TA98 and TA100 demonstrated the presence of the Pkm101 plasmid R-factor. All WP2 *uvr*A cultures demonstrated the deletion in the *uvr*A gene. All cultures demonstrated the characteristic mean number of spontaneous revertants in the vehicle controls as follows: TA98, 10-50; TA100, 80-240; TA1535, 5-45; TA1537, 3-21, WP2*uvr*A, 10-60.

VII. Conclusion

All criteria for a valid study were mete as described in the protocol. The results of the Bacterial Reverse Mutation Assay indicate that under the conditions of this assay, the test article solution was considered to be Non-Mutagenic to *Salmonella typhimurium* tester strains TA98, TA100, TA1537, TA1535 and *Escherichia coli* WP2*uvr*A.The negative and positive controls performed as anticipated. The results of this study should be evaluated in conjunction with other required tests as listed in ISO 100993, Part 3: Tests for Genotoxicity, Carcinogenicity, and Reproductive Toxicology.

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Appendix 2:

Bacterial Mutation Assay
Plate Incorporation Assay Results

	Concentration µg per Plate	***TA98***		
		Revertants per plate (CFU)		Mean
Test Solution w/ S9	5000	26	24	25
	1500	22	22	22
	500	36	35	36
	150	42	40	41
	50	45	55	50
	15	46	33	40
	5.0	52	46	49
	1.5	49	51	50
Test Solution w/o S9	5000	63	52	58
	1500	48	55	52
	500	73	65	69
	150	65	68	67
	50	72	56	64
	15	58	66	62
	5.0	61	55	58
	1.5	69	50	60
DI Water w/S9		54	58	56
DI Water w/o S9		56	61	59
2-aminoanthracen w/ S9		301	322	312
2-nitrofluorene w/o S9		210	351	281
Historical Count Positive w/S9		**43-1893**		
Historical Count Positive w/o S9		**39-1871**		
Historical Count Negative w/S9		**4-69**		
Historical Count Negative w/o S9		**3-59**		

*CFU = Colony Forming Units
*Mean = Average of duplicate plates

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	Concentration µg per Plate	***TA100***		
		Revertants per plate (CFU)		Mean
Test Solution w/ S9	5000	116	121	119
	1500	132	110	121
	500	145	136	141
	150	150	168	159
	50	163	122	143
	15	148	165	157
	5.0	155	116	136
	1.5	150	140	145
Test Solution w/o S9	5000	92	85	89
	1500	115	123	119
	500	125	138	132
	150	149	139	144
	50	156	112	134
	15	170	162	166
	5.0	163	152	158
	1.5	178	141	160
DI Water w/S9		120	148	134
DI Water w/o S9		124	68	96
2-aminoanthracen w/ S9		630	540	585
Sodium azide w/o S9		840	1104	972
Historical Count Positive w/S9		**224-3206**		
Historical Count Positive w/o S9		**226-1837**		
Historical Count Negative w/S9		**55-268**		
Historical Count Negative w/o S9		**47-250**		

*CFU = Colony Forming Units
*Mean = Average of duplicate plates

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	Concentration µg per Plate	***TA1537***		
		Revertants per plate (CFU)		Mean
Test Solution w/ S9	5000	13	18	16
	1500	12	15	14
	500	6	12	9
	150	12	16	15
	50	21	20	21
	15	15	18	17
	5.0	19	19	19
	1.5	10	21	16
Test Solution w/o S9	5000	4	8	6
	1500	6	6	6
	500	15	14	15
	150	13	19	16
	50	10	12	11
	15	13	20	17
	5.0	10	11	11
	1.5	14	18	16
DI Water w/S9		18	33	26
DI Water w/o S9		20	5	13
2-aminoanthracen w/ S9		150	136	143
2-aminoacridine w/o S9		210	202	206
Historical Count Positive w/S9		**13-1934**		
Historical Count Positive w/o S9		**17-4814**		
Historical Count Negative w/S9		**0-41**		
Historical Count Negative w/o S9		**0-29**		

*CFU = Colony Forming Units
*Mean = Average of duplicate plates

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	Concentration µg per Plate	***TA1535***		
		Revertants per plate (CFU)		Mean
Test Solution w/ S9	5000	16	19	18
	1500	22	21	22
	500	15	16	16
	150	26	21	24
	50	13	11	12
	15	20	15	18
	5.0	23	23	23
	1.5	15	19	17
Test Solution w/o S9	5000	11	17	14
	1500	13	15	14
	500	12	15	14
	150	15	21	18
	50	22	19	21
	15	21	24	23
	5.0	14	15	15
	1.5	20	19	20
DI Water w/S9		18	21	20
DI Water w/o S9		34	20	27
2-aminoanthracen w/ S9		231	304	268
Sodium azide w/o S9		616	632	624
Historical Count Positive w/S9		**22-1216**		
Historical Count Positive w/o S9		**47-1409**		
Historical Count Negative w/S9		**1-50**		
Historical Count Negative w/o S9		**1-45**		

*CFU = Colony Forming Units
*Mean = Average of duplicate plates

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	Concentration µg per Plate	***WP2uvrA***		
		Revertants per plate (CFU)		Mean
Test Solution w/ S9	5000	33	42	38
	1500	20	26	23
	500	30	26	28
	150	38	35	37
	50	42	40	41
	15	33	26	30
	5.0	45	36	41
	1.5	48	41	45
Test Solution w/o S9	5000	55	63	59
	1500	63	50	57
	500	57	48	53
	150	43	46	45
	50	38	65	52
	15	40	36	38
	5.0	50	53	52
	1.5	49	66	58
DI Water w/S9		67	55	61
DI Water w/o S9		49	59	54
2-aminoanthracen w/ S9		274	263	269
Methylmethanesulfonate w/o S9		310	324	317
Historical Count Positive w/S9		**44-1118**		
Historical Count Positive w/o S9		**42-1796**		
Historical Count Negative w/S9		**8-80**		
Historical Count Negative w/o S9		**8-84**		

*CFU = Colony Forming Units
*Mean = Average of duplicate plates

ACTIVE MICRO TECHNOLOGIES

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Cellular Viability Assay Analysis

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10201

Lot #: N220927E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
Cellular Viability Assay

Introduction

The cellular viability assay is useful for quantitatively measuring cell-mediated cytotoxicity, cell proliferation and mitochondrial metabolic activity. Increased metabolism in a cell indicates ample cellular respiration and adenosine triphosphate (ATP) production. ATP is the molecular energy of cells and is required in basic cell function and signal transduction. A decrease in ATP levels indicates cytotoxicity and decreased cell function while an increase in ATP levels indicates healthy cells.

The cellular viability assay was conducted to assess the ability of **FortiCulture Coconut** to increase cellular metabolic activity in cultured dermal fibroblasts.

Assay Principle

The assay utilizes a nonfluorescent dye, resazurin, which is converted to a fluorescent dye, resorufin, in response to chemical reduction of growth medium from cell growth and by respiring mitochondria. Healthy cells that are in a proliferative state will be able to easily convert resazurin into resorufin without harming the cells. This method is a more sensitive assay than other commonly used mitochondrial reductase dyes such as MTT. An increase in the signal generated by resazurin-conversion is indicative of a proliferative cellular state.

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Materials

A.	**Kit:**	PrestoBlue™ Cell Viability Reagent (Invitrogen; A13261)*
B.	**Incubation Conditions:**	37°C at 5% CO_2 and 95% relative humidity
C.	**Equipment:**	Forma humidified incubator; ESCO biosafety laminar flow hood; Light microscope; Pipettes
D.	**Cell Line:**	Normal Human Dermal Fibroblasts (ATCC; PCS-201-012)*
E.	**Media/Buffers:**	Fibroblast Basal Medium (ATCC; PCS-201-030)*; Fibroblast Growth Kit (ATCC; PCS-201-041)*
F.	**Culture Plate:**	Falcon flat bottom 96-well tissue culture treated plates*
G.	**Reagents:**	PrestoBlue™ reagent (10X)*
H.	**Other:**	Sterile disposable pipette tips

**Or suitable alternatives, subject to change without notice based off vendor availability*

Methods

Human dermal fibroblasts were seeded into 96-well tissue culture plates and allowed to grow to confluency in complete serum-free media. A dilution was performed resulting in **FortiCulture Coconut** concentrations of 0.01%, 0.02%, 0.05%, and 0.1% in complete serum-free media and incubated with fibroblasts for 24 hours.

Ten microliters of viability reagent were added to 90 µL of cell culture media in culture wells and a fluorometric measurement was taken at 560 nm for excitation and 590 nm for emission.

Assays were repeated three separate times with each sample run in duplicate. Duplicates for each replicate were averaged, and the average of all three experiments is displayed. Data was analyzed using a one-way ANOVA with statistical significance accepted at $p \leq 0.05$. Cellular metabolism results are shown as mean fluorescence units (MFU) and expressed as percentage change, calculated by the below equation:

$$Percent\ Change\ (\%) = \frac{MFU_{Sample} - MFU_{Control}}{MFU_{Control}} \times 100$$

Results

The data obtained from this study met criteria for a valid assay and the control performed as anticipated.

FortiCulture Coconut did not exhibit negative effects on cell metabolism.

ACTIVE MICRO TECHNOLOGIES

Cellular Viability Assay Analysis

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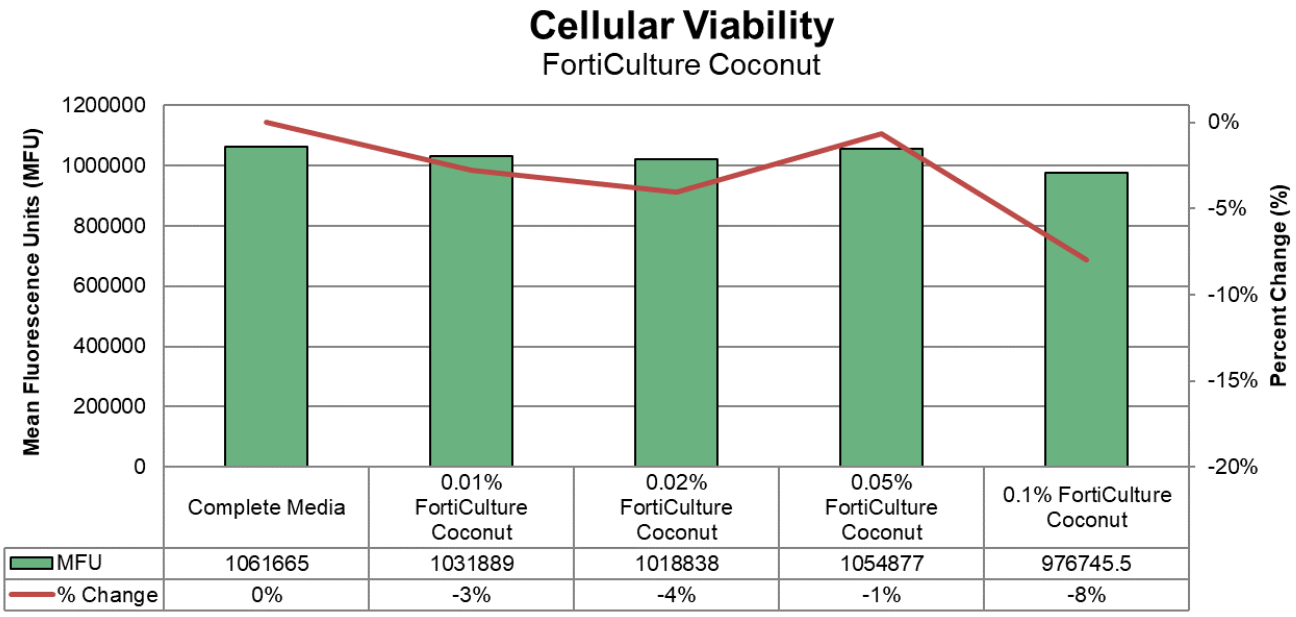

Figure 1: Cellular Metabolism of **FortiCulture Coconut-**Treated Fibroblasts

Table 1: P-values from one-way ANOVA Statistical Analysis Compared to Complete Media

	0.01% **FortiCulture Coconut**	0.02% **FortiCulture Coconut**	0.5% **FortiCulture Coconut**	0.1% **FortiCulture Coconut**
P-value	0.215	0.283	0.643	0.105

Discussion

In this study, **FortiCulture Coconut** was tested to evaluate its effects on the viability of normal human dermal fibroblasts. At concentrations of 0.01%, 0.02%, 0.05%, and 0.1%, **FortiCulture Coconut**, nor the preservatives contained therein exhibited any inhibition of cell viability.

Please note that when interpreting *in vitro* studies, a 0.1% concentration is comparable to a 10% dose in application. This high dosage can account for slightly decreased viability and efficacy *in vitro* and is included for comparison purposes.

It can therefore be concluded that at the suggested use concentrations **FortiCulture Coconut** is not cytotoxic.

ACTIVE MICRO TECHNOLOGIES

Dermal and Ocular Irritation Tests

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Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10485

Lot #: N230508E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
In Vitro EpiDerm™ Dermal Irritation Test (EPI-200-SIT)
EpiOcular™ Eye Irritation Test (OCL-200-EIT)

SUMMARY

In vitro dermal and ocular irritation studies were conducted to evaluate whether **FortiCulture Coconut** would induce dermal or ocular irritation in the EpiDerm™ and EpiOcular™ model assays.

The product was tested according to the manufacture's protocol. The test article solution was found to be a **non-irritant**. Reconstructed human epidermis and cornea epithelial model were incubated in growth media overnight to allow for tissue equilibration after shipping from MatTek Corporation, Ashland, MA. Test substances were applied to the tissue inserts and incubated for 60 minutes for liquid and solid substances in the EpiDerm™ assay and 30 minutes for liquid substances and 90 minutes for solid substances in the EpiOcular™ assay at 37°C, 5% CO_2, and 95% relative humidity (RH). Tissue inserts were thoroughly washed and transferred to fresh plates with growth media. After post substance dosing incubation is complete, the cell viability test begins. Cell viability is measured by dehydrogenase conversion of MTT [(*3-4,5-dimethyl thiazole 2-yl*)], present in the cell mitochondria, into blue formazan salt that is measured after extraction from the tissue. The irritation potential of the test chemical is dictated by the reduction in tissue viability of exposed tissues compared to the negative control.

Under the conditions of this assay, the test article was considered to be **non-irritating**. The negative and positive controls performed as anticipated.

I. Introduction

A. Purpose

In vitro dermal and ocular irritation studies were conducted to evaluate whether a test article would induce dermal or ocular irritation in the EpiDerm™ and EpiOcular™ model assays. MatTek Corporation's reconstructed human epidermal and human ocular models are becoming a standard in determining the irritancy potential of test substances. They are able to discriminate between irritants and non-irritants. The EpiDerm™ assay has accuracy for the prediction of UN GHS R38 skin irritating and no-label (non-skin irritating) test substances. The EpiOcular™ assay can differentiate chemicals that have been classified as R36 or R41 from the EU classifications based on Dangerous Substances Directive (DSD) or between the UN GHS Cat 1 and Cat 2 classifications.

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II. Materials

A. Incubation Conditions:	37°C at 5% CO_2 and 95% relative humidity
B. Equipment:	Forma humidified incubator, ESCO biosafety laminar flow hood, Synergy HT Microplate reader; Pipettes
C. Media/Buffers:	DMEM based medium; DPBS; sterile deionized H_2O
D. Preparation:	Pre-incubate (37°C) tissue inserts in assay medium; Place assay medium and MTT diluent at 4°C, MTT concentrate at -20°C, and record lot numbers of kit components
E. Tissue Culture Plates:	Falcon flat bottom 96-well, 24-well, 12-well, and 6-well tissue culture plates
F. Reagents:	MTT (1.0mg/mL); Extraction Solution (Isopropanol); SDS (5%); Methyl Acetate
G. Other:	Nylon Mesh Circles (EPI-MESH); Cotton tip swabs; 1mL tuberculin syringes; Ted Pella micro-spatula; 220mL specimen containers; sterile disposable pipette tips; Parafilm

III. Test Assay

A. Test System

The reconstructed human epidermal model, EpiDerm™, and cornea epithelial model, EpiOcular™, consist of normal human-derived epidermal keratinocytes which have been cultured to form a multilayer, highly differentiated model of the human epidermis and cornea epithelium. These models consist of organized basal, spinous, and granular layers, and the EpiDerm™ systems also contains a multilayer stratum corneum containing intercellular lamellar lipid layers that the EpiOcular™ system is lacking. Both the EpiDerm™ and EpiOcular™ tissues are cultured on specially prepared cell culture inserts.

B. Negative Control

Sterile DPBS and sterile deionized water are used as negative controls for the EpiDerm™ and EpiOcular™ assays, respectfully.

C. Positive Control

Known dermal and eye irritants, 5% SDS solution and Methyl Acetate, were used as positive controls for the EpiDerm™ and EpiOcular™ assays, respectfully.

D. Data Interpretation Procedure

a. EpiDerm™

An irritant is predicted if the mean relative tissue viability of the 3 tissues exposed to the test substance is reduced by 50% of the mean viability of the negative controls and a non-irritant's viability is > 50%.

b. EpiOcular™

An irritant is predicted if the mean relative tissue viability of the 2 tissues exposed to the test substance is reduced by 60% of the mean viability of the negative controls and a non-irritant's viability is > 40%.

IV. Method

A. Tissue Conditioning

Upon MatTek kit arrival at Active Micro Technologies, LLC the tissue inserts are removed from their shipping medium and transferred into fresh media and tissue culture plates and incubated at 37°C at 5% CO_2 and 95% relative humidity for 60 minutes. After those 60 minutes the inserts are transferred into fresh media and tissue culture plates and incubated at 37°C at 5% CO_2 and 95% relative humidity for an additional 18 to 21 hours.

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B. Test Substance Exposure

a. EpiDerm™

30µL (liquid) or 25mg (solid) of the undiluted test substance is applied to 3 tissue inserts and allowed to incubate for 60 minutes in a humidified incubator (37°C, 5% CO_2, 95% RH).

b. EpiOcular™

Each tissue is dosed with 20µL DPBS prior to test substance dosing. 50µL (liquid) or 50mg (solid) of the undiluted test substance is applied to 2 tissue inserts and allowed to incubate for 90 minutes in a humidified incubator (37°C, 5% CO_2, 95% RH).

C. Tissue Washing and Post Incubation

a. EpiDerm™

All tissue inserts are washed with DPBS, dried with cotton tipped swab, and transferred to fresh media and culture plates. After 24 hours the inserts are again transferred into fresh media and culture plates for an additional 18 to 20 hours.

b. EpiOcular™

Tissue inserts are washed with DPBS and immediately transferred into 5mL of assay medium for 12 to 14 minutes. After this soak the inserts are transferred into fresh media and tissue culture plates for 120 minutes for liquid substances and 18 hours for solid substances.

D. MTT Assay

Tissue inserts are transferred into 300µL MTT media in pre-filled plates and incubated for 3 hours at 37°C, 5% CO_2, and 95% RH. Inserts are then removed from the MTT medium and placed in 2mL of the extraction solution. The plate is sealed and incubated at room temperature in the dark for 24 hours. After extraction is complete the tissue inserts are pierced with forceps and 2 x 200µL aliquots of the blue formazan solution is transferred into a 96 well plate for Optical Density reading. The spectrophotometer reads the 96-well plate using a wavelength of 570 nm.

V. Acceptance Criterion

A. Negative Control

The results of this assay are acceptable if the mean negative control Optical Density (OD_{570}) is ≥ 1.0 and ≤ 2.5 (EpiDerm™) or ≥ 1.0 and ≤ 2.3 (EpiOcular™).

B. Positive Control

a. EpiDerm™

The assay meets the acceptance criterion if the mean viability of positive control tissues expressed as a % of the negative control is ≤ 20%.

b. EpiOcular™

The assay meets the acceptance criterion if the mean viability of positive control tissues is < 60% of control viability.

C. Standard Deviation

Since each irritancy potential is predicted from the mean viability of 3 tissues for EpiDerm™ and 2 tissues for EpiOcular™, the variability of the replicates should be < 18% for EpiDerm™ and < 20% EpiOcular™.

VI. Results

A. Tissue Characteristics

The tissue inserts included in the MatTek EpiDerm™ and EpiOcular™ assay kits were in good condition, intact, and viable.

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B. Tissue Viability Assay
The results are summarized in Figures 1 and 2. In no case was the tissue viability ≤ 50% for EpiDerm™ or ≤ 60% for EpiOcular™ in the presence of the test substance. The negative control mean exhibited acceptable relative tissue viability while the positive control exhibited substantial loss of tissue viability and cell death.

C. Test Validity
The data obtained from this study met criteria for a valid assay.

VII. Conclusion
Under the conditions of this assay, the test article substance was considered to be non-irritating. The negative and positive controls performed as anticipated.

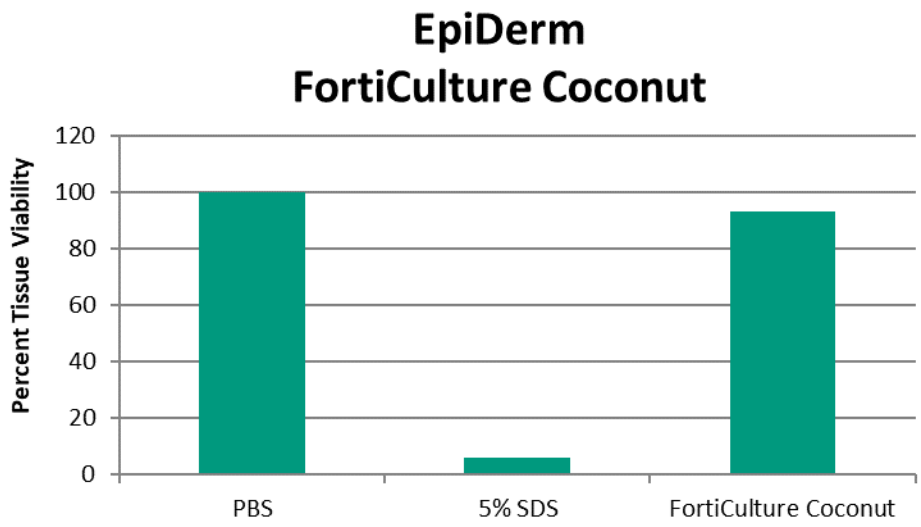

Figure 1: EpiDerm tissue viability

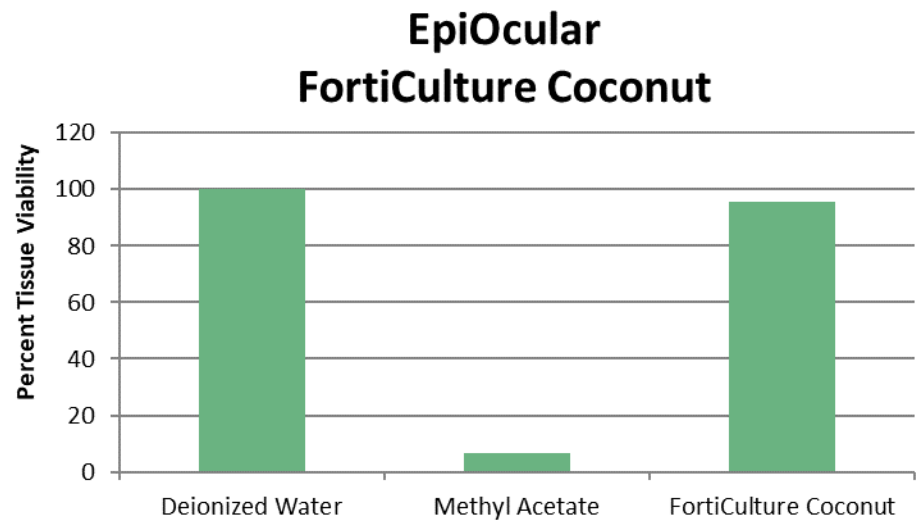

Figure 2: EpiOcular tissue viability

OECD 201 Freshwater Alga Growth Inhibition Test

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(704) 276-7100 • Fax (704) 276-7101

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10488

Lot #: N230508E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Daniel Shill*

Test Performed:
OECD 201
Freshwater Alga Growth Inhibition Test

Introduction

The purpose of the present study is to determine the toxicity of **FortiCulture Coconut** by exposing the exponentially growing test organism *Pseudokirchneriella subcapitata* to the test substance for 72 hours and measuring the growth and growth inhibition through cell counting against the control. The response is evaluated as a function of the exposure concentration in comparison with the average growth of replicate, unexposed control cultures.

OECD Guideline 201 on "Fresh Alga and Cyanobacteria, Growth Inhibition Test", adopted in 1984, extended the guideline to include additional species and update it to meet the requirements for hazard assessment and classification of chemicals in 2006.

Assay Principle

Pseudokirchneriella subcapitata, are exposed to the test substance at a range of concentrations for a period of 72 hours. The cultures are allowed unrestricted exponential growth under nutrient sufficient conditions and continuous light for a sufficient period of time to measure reduction of the specific growth rate. Growth and growth inhibition are quantified from measurements of the algal biomass as a function of time. The test endpoint is inhibition of growth, expressed as the logarithmic increase in biomass during the 72 hour exposure period. The results are analyzed in order to calculate the EC_{10} and EC_{20} at 72 hours. The response is evaluated as a function of the exposure concentration in comparison with the average growth of replicate, unexposed control cultures.

A reliable analytical method for the quantification of the substance in the test solutions with reported recovery efficiency and limit of determination should be available. A reference substance may be tested for EC_{50} as a means of assuring that the test conditions are reliable.

Analysis of the concentration of the test substance at the start and end of the test of a low and high test concentration around the expected EC_{50} may be sufficient where it is likely that exposures concentrations will vary less than 20% from the nominal values during the test.

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Materials

- Glass Flasks with air-permeable stopper
- Automated Pipette
- pH Meter
- Temperature Control Apparatus
- Microscope with counting chamber
- *Pseudokirchneriella subcapitata* (ATCC 22662)
- Gorham's medium for algae (ATCC MD-0625)

Methods

Test Conditions

- Incoculum Culture
  - Inoculum culture is incubated under the same conditions as the test cultures for 2-4 days allowing for exponential growth to prevail before the start of the test. This is done to ensure that growth is within the normal range for the test strain under the culturing conditions.
- Initial Biomass
  - The initial biomass in the test cultures must be the same in all test cultures and sufficiently low to allow exponential growth throughout the incubation period without risk of nutrient depletion. The initial biomass should not exceed 0.5 mg/L as dry weight.
- Exposure Period
  - 72 hours
- Number of Test Organisms
  - *Pseudokirchneriella subcapitata* 5 x 10^{3-4} cells/ml
- Test Concentration
  - Adopt a concentration range of at least 5 concentrations, causing a range of 5-75% inhibition of algal growth rate expressed as E_rC_x
- Culture Method
  - Illumination: Continuous uniform fluorescent illumination
  - Temperature: The temperature is between 21°C to 24°C
  - pH: pH of the control medium should not increase be more than 1.5 units during test

Measurement of Test Substance Concentrations

- Measurement of biomass is done by manual cell counting by microscope.
- Algal biomass in each flask is determined daily during test period.
- At the beginning and end of exposure, measure test substance concentrations at the lowest and highest test concentration groups.
  - For volatile or adsorptive substances, additional measurements are recommended at 24 hours intervals during exposure period.

Observation

- Microscopic observation should be performed to verify a normal and healthy appearance of the inoculum culture and to observe any abnormal appearance of the algae at the end of the test.

Test Condition Measurements

- Measure pH in the control and at the highest test concentration at the beginning and end of the exposure period.
- Water temperature should be measured at the beginning and end of the exposure period.

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Data and Reporting

I. Data
   a. Tabulate the estimated biomass concentration in test cultures and controls together with the concentrations of test materials and the times of measurement, recorded with a resolution of at least whole hours, to produce plots of growth curves.
   b. For each response variable to be analyzed, use the concentration-response relationship to calculate point estimates of EC_x values. Recent scientific developments have led to a recommendation of abandoning the concept of NOEC and replacing it with regression based point estimates EC_x, specifically EC_{10} and EC_{20}.

II. Test Report
   a. The test report must include the following:
      i. Test substance:
         1. Physical nature and relevant physical-chemical properties
         2. Chemical identification data, including purity
      ii. Test species:
         1. Source and species of *Pseudokirchneriella subcapitata*, supplier of source (if known), and the culture conditions (including source, kind and amount of food, feeding frequency)
      iii. Test conditions:
         1. Description of test vessels: type and volume of vessels, volume of solution, density of *Pseudokirchneriella subcapitata* per test vessel, number of test vessels (replicates) per concentration
         2. Methods of preparation of stock and test solutions including the use of any solvent or dispersants, concentrations used
         3. Details of dilution water: source and water quality characteristics (pH, hardness, Ca/Mg ratio, Na/K ratio, alkalinity, conductivity, etc); composition of reconstituted water if used
         4. Incubation conditions: temperature, light intensity and periodicity, pH, etc.
      iv. Results:
         1. The nominal test concentrations and the result of all analyses to determine the concentration of the test substance in the test vessels; the recovery efficiency of the method and the limit of determination should also be reported
         2. All physical-chemical measurements of temperature and pH made during the test
         3. The EC_{10} and EC_{20} at 72 hours for percent inhibition with confidence intervals and graphs of the fitted model used for calculation, the slopes of the dose-response curves and their standard error; statistical procedures used for determination of EC_{10} and EC_{20}.

$$Percent\ (\%)Inhibition = \frac{\mu c - \mu T}{\mu c} \times 100$$

μ_c: mean value for average specific growth rate (μ) in the control group
μ_T: average specific growth rate for the treatment replicate

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Results

General Information:

Name of new chemical substance	**FortiCulture Coconut**		
INCI Nomenclature	Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate		
CAS number	7732-18-5 & 8001-31-8 & 8013-01-2		
Formulation Method	Fermentation, Extraction		
Molecular weight	330 Da		
Purity of the new chemical substance used for the test (%)	100%		
Lot number of the new chemical substance used for the test	N230508E		
Names and contents of impurities	N/A		
Solubility in water	Soluble		
Properties at room temperature	Clear to Slightly Hazy Colorless to Light Yellow Liquid, Characteristic Odor		
Stability	Stable Under Normal Conditions		
Solubility in solvents, etc.	**Solvent**	**Solubility**	**Stability in solvent**
	N/A	N/A	N/A

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OECD 201 Freshwater Alga Growth Inhibition Test

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Test Materials and Methods:

Items			**Contents**
Test Organisms	Species		*Pseudokirchneriella subcapitata*
	Source		ATCC
	Reference substance (EC_{50})		3,5-dichlorophenol
Culture	Kind of Medium		Gorham's Medium for Algae
	Conditions (Temperature)		22°C ± 2°C
Test Conditions	Test Vessel		Glass
	Material Water	Kind	Deionized
		Hardness	250 mg/L
		pH	7.4
	Date of Exposure		05/22/2023
	Test Concentrations		200, 89.4, 42.3, 19.2, 7.8 mg/L
	Number of organisms		5 x 103-4 cells/ml
	Number of Replicates	Exposure Group	4
		Control Group	4
	Test Solution Volume		5 mL
	Vehicle	Use or Not	N/A
		Kind	N/A
		Concentration	N/A
		Number of Replicates	N/A
	Photoperiod		Continuous

Test Results:

Items		**Contents**
Toxicity Value	Percent Inhibition EC_{10} and EC_{20}	103.68 mg/L and 184.24 mg/L
Exposure Concentrations Used for Calculation	Nominal Values	200, 89.4, 42.3, 19.2, 7.8 mg/L
Remarks		Not harmful to aquatic organisms

Discussion

After 72 hours, the percent inhibition for **FortiCulture Coconut** was determined to be 103.68 mg/L EC_{10} and 184.24 mg/L EC_{20}. The conditions of OECD guideline 201 for the validity of the test were adhered to, this product is not classified and therefore not harmful to aquatic organisms.

OECD 301B Ready Biodegradability Assay

107 Technology Drive • Lincolnton, NC 28092
(704) 276-7100 • Fax (704) 276-7101

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10486

Lot #: N230508E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
OECD 301 B
Ready Biodegradability: CO_2 Evolution (Modified Sturm Test)

Introduction

A study was conducted to assess the ready biodegradability of **FortiCulture Coconut** in an aerobic aqueous medium. In the OECD guideline 301 for ready biodegradability, six methods are provided as options. This report uses method B, CO_2 Evolution, also known as a Modified Sturm Test. This method was chosen based on the solubility, volatility, and adsorbing capabilities of the test sample.

Assay Principle

A solution or suspension of the test substance in a mineral medium is inoculated and incubated under aerobic conditions in the dark or in diffuse light. The amount of DOC (Dissolved Organic Carbon) in the test solution due to the inoculum should be kept as low as possible compared to the amount of organic carbon due to the test substance. Allowance is made for the endogenous activity of the inoculum by running parallel blanks with inoculum but without test substance. A reference compound is run in parallel to check the procedures' operation.

In general, degradation is followed by the determination of parameters such as DOC, carbon dioxide production, and oxygen uptake. Measurements are taken at sufficiently frequent intervals to allow the identification of the beginning and end of biodegradation.

Normally this test lasts for 28 days, but it may be ended before that time if the biodegradation curve reaches a plateau for at least three determinations. Tests may also be prolonged beyond 28 days when the curve shows that biodegradation has started but the plateau has not yet been reached. In such cases the test substance would not be classified as readily biodegradable.

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The pass levels for ready biodegradability are 70% removal of DOC and 60% of ThOD (Theoretical Oxygen Demand) or $ThCO_2$ (Theoretical Carbon Dioxide) production for respirometric methods. They are lower in the respirometric methods since, as some of the carbon from the test chemical is incorporated into new cells, the percentage of CO_2 produced is lower than the percentage of carbon being used. These pass values have to be reached in a 10-day window within the 28-day period of the test. The 10-day window begins when the degree of biodegradation has reached 10% DOC, ThOD, or $ThCO_2$ and must end before day 28 of the test. Test substances which reach the pass levels after the 28-day period are not deemed to be readily biodegradable.

In order to check the procedure, reference compounds which meet the criteria for ready biodegradability are tested by setting up an appropriate vessel in parallel as part of normal test runs. Suitable compounds are freshly distilled aniline, sodium acetate, and sodium benzoate. These compounds all degrade in this method even when no inoculum is deliberately added.

Because of the nature of biodegradation and of the mixed bacterial populations used as inocula, determinations should be carried out at least in duplicate. It is usually found that the larger the concentration of microorganisms initially added to the test medium, the smaller the variation between replicates.

Materials

- Water
  - Deionized or distilled, free from inhibitory concentrations of toxic substances
  - Must contain no more than 10% of the organic carbon content introduced by the test material
  - Use only one batch of water for each series of tests
- Mineral media
  - To prepare the mineral medium, mix 10 mL of solution A with 800 mL water. Then add 1 mL each of solutions B, C, and D and make up to 1 liter with water.
  - Solution A (Dissolve in water and make up to 1 liter; pH 7.4)
    - Potassium dihydrogen orthophosphate, KH_2PO.....8.5g
    - Dipotassium hydrogen orthophosphate, K_2HPO_4.....21.8g
    - Disodium hydrogen orthophosphate dehydrate, $Na_2HPO_4.2H_2O$.....33.4g
    - Ammonium chloride, NH_4Cl.....0.5g
  - Solution B (Dissolve in water and make up to 1 liter)
    - Calcium chloride, anhydrous, $CaCl_2$.....27.50g
      Or
    - Calcium chloride dehydrate, $CaCl_2.2H_2O$.....36.40g
  - Solution C (Dissolve in water and make up to 1 liter)
    - Magnesium sulphate heptahydrate, $MgSO_4.7H_2O$.....22.50g
  - Solution D (Dissolve in water and make up to 1 liter.)
    - Iron (III) chloride hexahydrate, $FeCl_3.6H_2O$.....0.25g
  - Flasks, 2-5 liters each, fitted with aeration tubes reaching nearly to the bottoms of the vessels and an outlet
  - Magnetic stirrers
  - Gas absorption bottles
  - Device for controlling and measuring air flow
  - Apparatus for carbon dioxide scrubbing, for preparation of air which is free from carbon dioxide; alternatively, a mixture of CO_2-free oxygen and CO_2-free nitrogen from gas cylinders in the correct proportions (20% O_2 : 80% N_2)
  - Device for determination of carbon dioxide, either titrimetrically or by some form of inorganic carbon analyzer

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- Stock solutions of test substances
  - When solubility of the substance exceeds 1 g/L, dissolve 1-10 g, as appropriate, of test or reference substance in water and make up to 1 liter. Otherwise, prepare stock solutions in mineral medium or add the chemical directly to the mineral medium, making sure it
- Inoculum
  - The inoculum may be derived from the following sources
    - Activated sludge
    - Sewage effluents
    - Surface waters
    - Soils
    - Or from a mixture of these.
  - Inoculum may be pre-conditioned to the experimental conditions, but not pre-adapted to the test substance. Pre-conditioning consists of aerating activated sludge in mineral medium or secondary effluent for 5-7 days at the test temperature. Pre-conditioning sometimes improves the precision of the test method by reducing blank values.

Methods

I. Preparation of flasks: As an example, the following volumes and weights indicate the values for 5-liter flasks containing 3 liters of suspension. If smaller volumes are used, modify the values accordingly.
   a. To each 5-liter flask, add 2,400 mL mineral medium.
   b. Add an appropriate volume of the prepared activated sludge to give a concentration of suspended solids of not more than 30 mg/L in the final 3 liters of inoculated mixture. Alternatively, first dilute the prepared sludge to give a suspension of 500-1000 mg/L in the mineral medium before adding an aliquot to the contents of the 5-liter flask to attain a concentration of 30 mg/L.
   c. Aerate these inoculated mixtures with CO_2-free air overnight to purge the system of carbon dioxide.
   d. Add the test material and reference compound, separately, as known volumes of stock solutions, to replicate flasks to yield concentrations, contributed by the added chemicals, of 10 – 20 mg DOC or TOC per liter. Leave some flasks without addition of chemicals as inoculum controls. Add poorly soluble test substances directly to the flasks on a weight or volume basis. Make up the volumes of suspensions in all flasks to 3 liters by the addition of mineral medium previously aerated with CO_2-free air.
   e. If required, use one flask to check the possible inhibitory effect of the test substance by adding both the test and reference substances at the same concentrations as present in the other flasks.
   f. If required, check whether the test substance is degraded abiotically by using a sterilized uninoculated solution of the chemical. Sterilize by the addition of a toxic substance at an appropriate concentration.
   g. If barium hydroxide is used, connect three absorption bottles, each containing 100 mL of 0.0125M barium hydroxide solution, in series to each 5-liter flask. The solution must be free of precipitated sulfate and carbonate and its strength must be determined immediately before use.
   h. If sodium hydroxide is used, connect two traps, the second acting as a control to demonstrate that all the carbon dioxide was absorbed in the first. Absorption bottles fitted with serum bottle closures are suitable. Add 200 mL 0.05M sodium hydroxide to each bottle. This is sufficient to absorb the total quantity of carbon dioxide evolved when the test substance is completely degraded.
   i. In a typical run, the following flasks are used:
      i. Flasks 1 & 2: containing test substance and inoculum (test suspension)
      ii. Flasks 3 & 4: containing only inoculum (inoculum blank)
      iii. Flask 5: containing reference compound and inoculum (procedure control)
      iv. Flask 6: containing test substance and sterilizing agent (abiotic sterile control)
      v. Flask 7: containing test substance, reference compound and inoculum (toxicity control)

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II. Start the test by bubbling CO_2-free air through the suspensions at a rate of 30-100 mL/minute.

III. CO_2 Determination
   a. It is mandatory to follow the CO_2 evolution from the test suspensions and inoculum blanks in parallel and it is advisable to do the same for the other test vessels.
   b. During the first ten days it is recommended that analyses of CO_2 should be made every second or third day and then at least every fifth day until the 28th day so that the 10-day window period can be identified. On the days of CO_2 measurement, disconnect the barium hydroxide absorber closest to the test vessel and titrate the hydroxide solution with 0.05M HCl using phenolphthalein as the indicator. Move the remaining absorbers one place closer to the test vessel and place a new absorber containing 100 mL fresh 0.0125M barium hydroxide at the far end of the series. Make titrations are needed (for example, when substantial precipitation is seen in the first trap and before any is evident in the second, or at least weekly). Alternatively, with NaOH as absorbent, withdraw a sample of the sodium hydroxide solution from the absorber nearest to the test vessel using a syringe. The sample volume needed will depend on the carbon analyzer used, but sampling should not significantly change the absorbent volume over the test period. Inject the sample into the IC part of the carbon analyzer for analysis of evolved carbon dioxide directly. Analyze the contents of the second trap only at the end of the test in order to correct for any carry-over of carbon dioxide.
   c. On the 28th day withdraw samples, optionally, for DOC and/or specific chemical analysis. Add 1 mL of concentrated hydrochloric acid to each test vessel and aerate them overnight to drive off the carbon dioxide present in the test suspensions. On day 29 make the last analysis of evolved carbon dioxide.

Data and Reporting

I. Treatment of Results
   a. Data from the test should be entered onto the attached data sheet.
   b. The amount of CO_2 produced is calculated from the amount of base remaining in the absorption bottle. When 0.0125M $Ba(OH)_2$ is used as the absorbent, the amount remaining is assessed by titrating with 0.05M HCl.
   c. Since 1 mmol of CO_2 is produced for every mol of $Ba(OH)_2$ reacted to $BaCl_2$ and 2 mmol of HCl are needed for the titration of the remaining $Ba(OH)_2$ and given that the molecular weight of CO_2 is 44 g, the weight of CO_2 produced (in mg) is calculated by:

$$\frac{0.05 \times (50 - mL\ HCl\ Titrated) \times 44}{2} = 1.1 \times (50 - mL\ HCl\ Titrated)$$

   Therefore, the factor to convert volume of HCl titrated to mg CO_2 produced is 1.1 in this case. Calculate the weights of CO_2 produced from the inoculum alone and from the inoculum plus test substance using the respective titration values. The difference is the weight of CO_2 produced from the test substance alone.

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d. The percentage biodegradation is calculated from:

$$\%\ Degradation = \frac{mg\ CO_2\ Produced}{ThCO_2 \times mg\ Test\ Substance\ Added} \times 100$$

Or

$$\%\ Degradation = \frac{mg\ CO_2\ Produced}{mg\ TOC\ Added\ in\ Test\ \times 3.67} \times 100$$

$$Where\ 3.67\ is\ the\ conversion\ factor\ \left(\frac{44}{12}\right)\ for\ carbon\ to\ carbon\ dioxide$$

e. When NaOH is used as the absorbent, calculate the amount of CO_2 produced after any time interval from the concentration of inorganic carbon and the volume of absorbent used. Calculate the percentage degradation from:

$$\%\ ThCO_2 = \frac{mg\ IC\ from\ Test\ Flask - mg\ IC\ from\ Blank}{mg\ TOC\ Added\ as\ Test\ Substances} \times 100$$

f. Display the course of degradation graphically and indicate the 10-day window. Calculate and report the percentage removal achieved at the plateau, at the end of the test, and/or at the end of the 10-day window, whichever is appropriate.
g. When appropriate, calculate DOC removals using the equation given in 301 A paragraph 27.
h. When an abiotic control is used, calculate the percentage abiotic degradation by:

$$\%\ Abiotic\ Degradation = \frac{CO_2\ Produced\ by\ Sterile\ Flask\ After\ 28\ Days\ (mg)}{ThCO_2\ (mg)} \times 100$$

Validity of Tests

The IC content of the test substance suspension in the mineral medium at the beginning of the test must be less than 5% of the TC, and the total CO_2 evolution in the inoculum blank at the end of the test should not normally exceed 40 mg/L medium. If values greater than 70 mg CO_2/L are obtained, the data and experimental technique should be examined critically.

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OECD 301B Ready Biodegradability Assay

Data Sheet

Laboratory	Active Concepts Tissue Culture Laboratory		
Test Start Date	05/22/2023		
Test Substance	**Name**	**FortiCulture Coconut**	
	Stock Solution Concentration	2 g/L	
	Initial Concentration in Medium	20 mg/L	
Inoculum	**Source**	Activated Sludge	
	Treatment Given	Centrifugation	
	Pre-conditioning	N/A	
	Suspended Solids Concentration in Reaction Mixture	4 mg/L	
Reference Material	Sodium Benzoate	**Concentration**	20 mg/L
CO_2 Production and Degradability	**Method**	**$Ba(OH)_2$**	0.0125M
		NaOH	N/A
		Other	N/A
Total Contact Time	28 Days		
Total CO_2 Evolved Measurements	**Days**	2, 4, 11, 17, 23, 28	
Degradation Over Time	93.9% and 94.4% after 28 days		
Remarks	Test material was readily biodegradable		
Conclusion	This test met the criteria for a valid assay		

Discussion

Based on the testing conducted in accordance with the specified test method, **FortiCulture Coconut** achieved 94.2% biodegradation after 28 days of testing. The product met method requirements for the Readily Biodegradable classification.

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OECD TG 442C: *In Chemico* Skin Sensitization

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10487

Lot #: N230508E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
OECD TG 442C: *In Chemico* Skin Sensitization
Direct Peptide Reactivity Assay (DPRA)

Introduction

A skin sensitizer is a substance that will lead to an allergic response following skin contact[1]. Haptenation is the covalent binding of a hapten, or low-molecular weight substance or chemical, to proteins in the skin. This is considered the prominent mechanism which defines a chemical as a sensitizer. Haptenation is described as a "molecular initiating event" in the OECD Adverse Outcome Pathway (AOP) for skin sensitization which summarizes the key events known to be involved in chemically-induced allergic contact dermatitis[2]. The direct peptide reactivity assay (DPRA) is designed to mimic the covalent binding of electrophilic chemicals to nucleophilic centers in skin proteins by quantifying the reactivity of chemicals towards the model synthetic peptides containing cysteine and lysine. The DPRA is able to distinguish sensitizers from non-sensitizer with 82% accuracy (sensitivity of 76%; specificity of 92%)[3].

This assay was conducted to determine skin sensitization hazard of **FortiCulture Coconut** in accordance with European Union Reference Laboratory for Alternatives to Animal Testing (EURL ECVAM) and OECD Test Guideline 442C.

Assay Principle

The DPRA is an *in chemico* method which addresses peptide reactivity by measuring depletion of synthetic heptapeptides containing either cysteine or lysine following 24 hours incubation with the test substance. The peptide is a custom material containing phenylalanine to aid in detection. Depletion of the peptide in the reaction mixture is measured by HPLC with gradient elution and UV detection at 220 nm. Cysteine and lysine peptide percent depletion values are then calculated and used in a prediction model which allows assigning the test chemical to one of four reactivity classes used to support the discrimination between sensitizers and non-sensitizers.

1. United Nations Economic Commission (UNECE) (2013) Global Harmonized System of Classification and Labelling of Chemicals (GHS) 5th Revised Edition
2. OECD (2012). The Adverse Outcome Pathway for Skin Sensitization Initiated by Covalent Binding to Proteins. Part 1: Scientific Evidence. Series on Testing and Assessment No. 168
3. EC EURL ECVAM (2012) Direct peptide reactivity assay (DPRA) validation study report; pp 1 -74.

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Materials

A.	**Equipment:**	HPLC-UV (Waters Breeze - Waters 2998 Photodiode Array Detector); Pipettes; Analytical balance
B.	**HPLC/Guard Columns:**	Agilent Zorbax SB-C18 2.1mm x 100mm x 3.5µm; Phenomenex Security Guard C18 4mm x 2mm
C.	**Chemicals:**	Trifluoroacetic acid; Ammonium acetate; Ammonium hydroxide; Acetonitrile; Cysteine peptide (Ac-RFAA**C**AA-COOH); Lysine peptide (Ac-RFAA**K**AA-COOH); Cinnamic aldehyde
D.	**Reagents/Buffers:**	Sodium phosphate buffer (100mM); Ammonium acetate buffer (100mM)
E.	**Other:**	Sterile disposable pipette tips

Methods

Solution Preparation:

- 0.667mM Cysteine Peptide in 100mM Phosphate Buffer (pH 7.5)
- 0.667mM Lysine Peptide in 100mM Ammonium Acetate Buffer (pH 10.2)
- 100mM Cinnamic Aldehyde in Acetonitrile
- 100mM* **FortiCulture Coconut** in Acetonitrile

*For mixtures and multi-constituent substances of known composition such as **FortiCulture Coconut**, a single purity should be determined by the sum of the proportion of its constituents (excluding water), and a single apparent molecular weight determined by considering the individual molecular weights of each component in the mixture (excluding water) and their individual proportions. The resulting purity and apparent molecular weight can then be used to calculate the weight of test chemical necessary to prepare a 100 mM solution.

Reference Controls:

- Reference Control A: For calibration curve accuracy
- Reference Control B: For peptide stability over analysis time of experiment
- Reference Control C: For verification that the solvent does not impact percent peptide depletion

Sample, Reference Control, and Co-Elution Control Preparation:

- Once these solutions have been made they should be incubated at room temperature, protected from light, for 24±2 hours before running HPLC analysis.
- Each chemical should be analyzed in triplicate.

1:10 Ratio, Cysteine Peptide 0.5mM Peptide, 5mM Test Chemical	1:50 Ratio, Lysine Peptide 0.5mM Peptide, 25mM Test Chemical
• 750µL Cysteine Peptide Solution (or 100mM Phosphate Buffer, pH 7.5, for Co-Elution Controls) • 200µL Acetonitrile • 50µL Test Chemical Solution (or Acetonitrile for Reference Controls)	• 750µL Lysine Peptide Solution (or 100mM Ammonium Acetate Buffer, pH 10.2, for Co-Elution Controls) • 250µL Test Chemical Solution (or Acetonitrile for Reference Controls)

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Calibration Curve:

- Standards are prepared in a solution of 20% Acetonitrile:Buffer
  - For the Cysteine peptide using the phosphate buffer, pH 7.5
  - For the Lysine peptide using the ammonium acetate buffer, pH 10.2

	Standard 1	Standard 2	Standard 3	Standard 4	Standard 5	Standard 6	Standard 7
mM Peptide	0.534	0.267	0.1335	0.0667	0.0334	0.0167	0.000

HPLC Analysis:

- HPLC-UV system should be equilibrated at 30°C with 50% Mobile Phase A (0.1% (v/v) trifluoroacetic acid in water) and 50% Mobile Phase B (0.085% (v/v) trifluoroacetic acid in acetonitrile) for 2 hours
- Absorbance is measured at 220nm
- Flow Conditions:

Time	Flow	%A	%B
0 minutes	0.35 mL/min	90	10
10 minutes	0.35 mL/min	75	25
11 minutes	0.35 mL/min	10	90
13 minutes	0.35 mL/min	10	90
13.5 minutes	0.35 mL/min	90	10
20 minutes	End Run		

Data and Reporting

Acceptance Criteria:

1. The following criteria must be met for a run to be considered valid:
   a. Standard calibration curve should have an $r^2 > 0.99$.
   b. Mean percent peptide depletion values of three replicates for the positive control cinnamic aldehyde should be between 60.8% and 100% for the cysteine peptide and between 40.2% and 69% for the lysine peptide and the maximum standard deviation should be <14.9 for the percent cysteine depletion and <11.6 for the percent lysine depletion.
   c. Mean peptide concentration of reference controls A should be 0.50±0.05mM and the coefficient of variable of the peptide peak areas for reference B and C in acetonitrile should be <15.0%.

2. The following criteria must be met for a test chemical's results to be considered valid:
   a. Maximum standard deviation should be <14.9 for percent cysteine depletion and <11.6 for percent lysine depletion.
   b. Mean peptide concentration of the three reference control C should be 0.50±0.05mM.

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Prediction Model:

Cysteine 1:10/Lysine 1:50 Prediction Model		
Mean of Cysteine and Lysine % Depletion	**Reactivity Class**	**Prediction**
0% < Mean % Depletion < 6.38%	Minimal Reactivity	Non-sensitizer
6.38% < Mean % Depletion < 22.62%	Low Reactivity	Sensitizer
22.62% < Mean % Depletion < 42.47%	Moderate Reactivity	Sensitizer
42.47% < Mean % Depletion < 100%	High Reactivity	Sensitizer

If co-elution occurs with the lysine peptide, than the cysteine 1:10 prediction model can be used:

Cysteine 1:10 Prediction Model		
Mean of Cysteine and Lysine % Depletion	**Reactivity Class**	**Prediction**
0% < Cys % Depletion < 13.89%	Minimal Reactivity	Non-sensitizer
13.89% < Cys % Depletion < 23.09%	Low Reactivity	Sensitizer
23.09% < Cys % Depletion < 98.24%	Moderate Reactivity	Sensitizer
98.24% < Cys % Depletion < 100%	High Reactivity	Sensitizer

Therefore the measured values of % depletion in the three separated runs for each peptide depletion assay include:

Cysteine 1:10/Lysine 1:50 Prediction Model		
Mean of Cysteine and Lysine % Depletion	**Reactivity Class**	**Prediction**
3.18	Minimal Reactivity	Non-sensitizer
3.15	Minimal Reactivity	Non-sensitizer
3.16	Minimal Reactivity	Non-sensitizer

Cysteine 1:10 Prediction Model		
Mean of Cysteine and Lysine % Depletion	**Reactivity Class**	**Prediction**
2.98	Minimal Reactivity	Non-sensitizer
3.01	Minimal Reactivity	Non-sensitizer
3.02	Minimal Reactivity	Non-sensitizer

Results and Discussion

The data obtained from this study met criteria for a valid assay and the controls performed as anticipated.

Percent peptide depletion is determined by the following equation:

$$Percent\ Peptide\ Depletion = \left[1 - \left(\frac{Peptide\ Peak\ Area\ in\ Replicate\ Injection}{Mean\ Peptide\ Peak\ Area\ in\ Reference\ Controls\ C}\right)\right] \times 100$$

Based on HPLC-UV analysis of **FortiCulture Coconut (M11001)** we can determine this product is not classified as a sensitizer and is not predicted to cause allergic contact dermatitis. The Mean Percent Depletion of Cysteine and Lysine was 3.08% causing minimal reactivity in the assay giving us the prediction of a non-sensitizer.

ACTIVE MICRO TECHNOLOGIES

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OECD TG 442D: *In Vitro* Skin Sensitization

Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: 10489

Lot #: N230508E

Sponsor: *Active Micro Technologies, LLC; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
OECD TG 442D: In Vitro Skin Sensitization
ARE-Nrf2 Luciferase Test Method

Introduction

Skin sensitization refers to an allergic response following skin contact with the tested chemical, as defined by the United Nations Globally Harmonized System of Classification and Labelling of Chemicals[1]. Substances are classified as skin sensitizers if there is evidence in humans that the substance can lead to sensitization by skin contact or positive results from appropriate tests, both *in vivo* and *in vitro*. Utilization of the KeratinoSens™ cell line allows for valid *in vitro* testing for skin sensitization.

This assay was conducted to determine skin sensitization potential of **FortiCulture Coconut** in accordance with the UN GHS.

Assay Principle

The ARE-Nrf2 luciferase test method addresses the induction of genes that are regulated by antioxidant response elements (ARE) by skin sensitizers. The Keap1-Nrf2-ARE pathways have been shown to be major regulator of cytoprotective responses to oxidative stress or electrophilic compounds. These pathways are also known to be involved in the cellular processes in skin sensitization. Small electrophilic substances such as skin sensitizers can act on the sensor protein Keap1 (Kelch-like ECH-associated protein 1), by covalent modification of its cysteine residue, resulting in its dissociation from the transcription factor Nrf2 (nuclear factor-erythroid 2-related factor 2). The dissociated Nrf2 can then activate ARE-dependent genes such as those coding for phase II detoxifying enzymes.

The skin sensitization assay utilizes the KeratinoSens™ method which uses an immortalized adherent human keratinocyte cell line (HaCaT cell line) that has been transfected with a selectable plasmid to quantify luciferase gene induction as a measure of activation of Keap1-Nrf2-antioxidant/electrophile response element (ARE). This test method has been validated by independent peer review by the EURL-ECVAM. The addition of a luciferin containing reagent to the cells will react with the luciferase produced in the cell resulting in luminescence which can be quantified with a luminometer.

1. United Nations (UN) (2013). Globally Harmonized System of Classification and Labelling of Chemicals (GHS), Fifth revised edition, UN New York and Geneva, 2013

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Materials

A. Incubation Conditions:	37°C at 5% CO_2 and 95% relative humidity (RH)
B. Equipment:	Humidified incubator; Biosafety laminar flow hood; Microplate Reader; Pipettes
C. Cell Line:	KeratinoSens™ by Givaudan Schweiz AG
D. Media/Buffers:	Dulbecco's Modified Eagle Medium (DMEM); Fetal Bovine Serum (FBS); Phosphate Buffered Saline (PBS); Geneticin
E. Culture Plate:	Flat bottom 96-well tissue culture treated plates
F. Reagents:	Dimethyl Sulfoxide (DMSO); Cinnamic Aldehyde; ONE-Glo Reagent; 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT); sodium lauryl sulfate (SLS)
G. Other:	Sterile disposable pipette tips; wash bottles

Methods

KeratinoSens™ were into seeded four 96-well tissue culture plates and allowed to grow to 80 – 90% confluency in DMEM containing 10% FBS and 500µg/mL G418 geneticin. Twelve test concentrations of **FortiCulture Coconut** were prepared in DMSO with a concentration range from 0.98 - 2000 µM. These 12 concentrations were assayed in triplicate in 2 independently performed experiments. The positive control was cinnamic aldehyde for which a series of 5 concentrations prepared in DMSO had final test concentrations of 4 – 64µM. The negative control was a 1% test concentration of DMSO.

24 hour post KeratinoSens™ seeding, the culture media was removed and replaced with fresh media containing 10% FBS without G418 geneticin. 50 µL of the above described test concentrations was added to the appropriate wells. The treated plates were then incubated for 48 hours at 37°C in the presence of 5% CO_2 and 95% relative humidity. After treatment incubation was complete the media was removed and the wells were washed with PBS 3 times.

One of the four plates was used for a cytotoxicity endpoint, where MTT was added to the wells and incubated for 4 hours at 37°C in the presence of 5% CO_2. SLS was then added to the wells and incubated overnight at room temperature. A spectrometer measured the absorbance at 570 nm. The absorbance values (optical density) were then used to determine the viability of each well by comparing the optical density of each test material treated well to that of the solvent control wells to determine the IC_{50} and IC_{30} values.

The remaining 3 plates were used in the luciferase induction endpoint of the assay. 100 µL of Promega's ONE-Glo Reagent was added to 100 µL of fresh media containing 10% FBS without geneticin. Cells were incubated for 5 minutes to induce cell lysis and release luciferin into the media. Plates were read with a luminometer and $EC_{1.5}$ and maximum response (I_{max}) values were obtained.

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Data and Reporting

Acceptance Criteria:

1. Gene induction obtained with the positive control, cinnamic aldehyde, should be statistically significant above the threshold of 1.5 in at least one of the tested concentrations (from 4 to 64 µM).
2. The EC1.5 value should be within two standard deviations of the historical mean and the average induction in the three replicates for cinnamic aldehyde at 64 µM should be between 2 and 8.
3. The average coefficient of variability of the luminescence reading for the negative (solvent) control DMSO should be below 20% in each experiment.

A KeratinoSens™ prediction is considered positive if the following conditions are met:

1. The Imax is higher than 1.5-fold and statistically significantly higher as compared to the solvent (negative) control
2. The cellular viability is higher than 70% at the lowest concentration with a gene induction above 1.5 fold (i.e., at the EC1.5 determining concentration)
3. The $EC_{1.5}$ value is less than 1000 µM (or < 200 µg/ml for test chemicals with no defined MW)
4. There is an apparent overall dose-response for luciferase induction

Results

Compound	Classification	$EC_{1.5}$ (µM)	IC_{50}	I_{max}
Cinnamic aldehyde	Sensitizer	19	289.19 µM	31.86
DMSO	Non-Sensitizer	No Induction	243.24 µM	0.18
FortiCulture Coconut	Non-Sensitizer	No Induction	> 1000 µM	0.31

Table 1: Overview of KeratinoSens™ Assay Results

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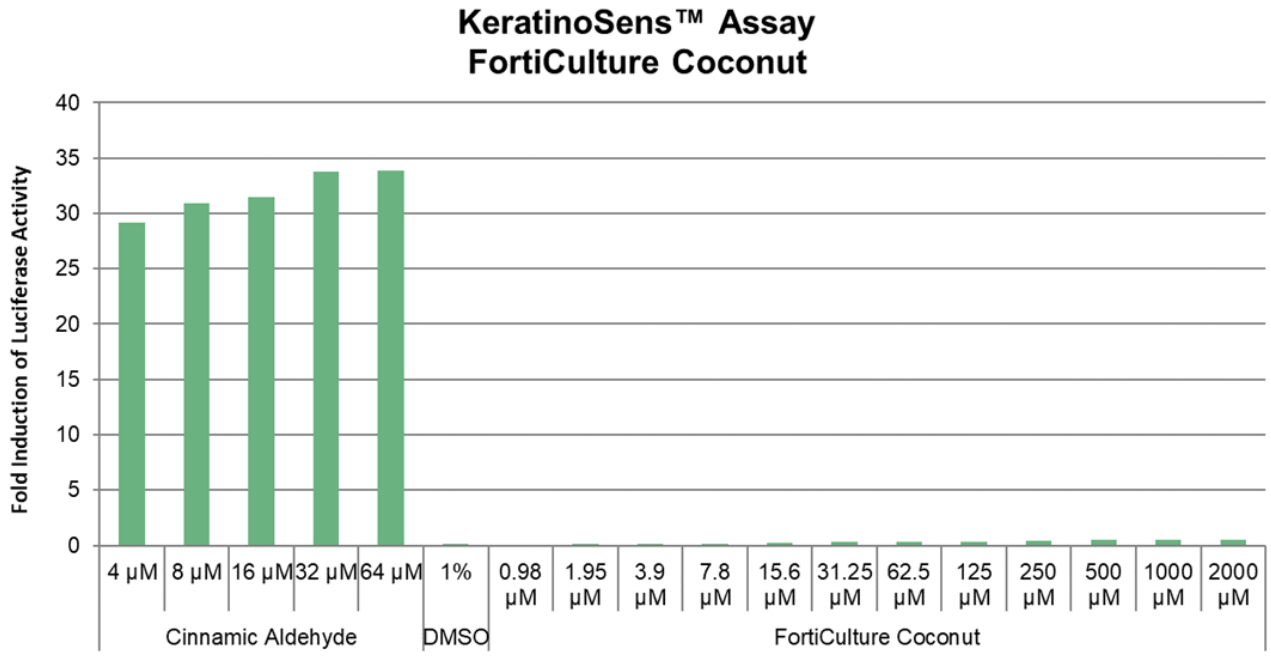

Figure 1: Fold Induction of Luciferase

Discussion

As shown in the results, **FortiCulture Coconut (M11001)** was not predicted to be a skin sensitizer based on the KeratinoSens™ ARE-Nrf2 Luciferase Test Method as there was not a significant increase in luciferase expression. It can be concluded that **FortiCulture Coconut** can be safely used in cosmetics and personal care products at typical use levels.

ACTIVE MICRO TECHNOLOGIES

Phototoxicity Assay Analysis

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Tradename: FortiCulture Coconut

Code: M11001

CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2

Test Request Form #: N230508E

Lot #: 10490

Sponsor: Active Micro Technologies, LLC*; 107 Technology Drive Lincolnton, NC 28092*
Study Director: *Maureen Drumwright*
Principle Investigator: *Hannah Duckett*

Test Performed:
In Vitro EpiDerm™ Model (EPI-200-SIT) Phototoxicity

SUMMARY

In vitro phototoxicity irritation studies were conducted to evaluate whether **FortiCulture Coconut** would induce phototoxic irritation in the EpiDerm™ model assay.

The product was tested according to the manufacturer's protocol. The test article solution was found to be a **non-photoirritant** at concentrations of 0.5%, 1.5%, 5.0% and 10.0%. Reconstructed human epidermis was incubated in growth media for one hour to allow for tissue equilibration after shipping from MatTek Corporation, Ashland, MA. Test substance was applied to the tissue inserts in four varying concentrations and incubated overnight at 37°C, 5% CO_2, and 95% relative humidity (RH). The following day, the appropriate tissue inserts were irradiated (UVA) for 60 minutes with 1.7 mW/cm^2 (=6 J/cm^2). After substance incubation, irradiation, and washing was completed, the cell viability test was conducted. Cell viability was measured by dehydrogenase conversion of MTT [(*3-4,5-dimethyl thiazole 2-yl*)], present in the cell mitochondria, into blue formazan salt that was measured after extraction from the tissue. The photoirritation potential of the test chemical was dictated by the reduction in tissue viability of UVA exposed tissues compared to non-UVA exposed tissues.

Under the conditions of this assay, the test article was considered to be **non-phototoxic** at concentrations of 0.5%, 1.5%, 5.0%, and 10.0%. The negative and positive controls performed as anticipated.

I. Introduction

A. Purpose

In vitro dermal phototoxicity study was conducted to evaluate whether a test article would induce photoirritation in the EpiDerm™ model assay. MatTek Corporation's reconstructed human epidermal model is becoming a standard in determining the phototoxicity potential of a test substance. This assay is able to discriminate between photoirritants and non-photoirritants at varying concentrations.

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II. Materials

A. Incubation Conditions:	37°C at 5% CO_2 and 95% relative humidity
B. Equipment:	Forma humidified incubator, ESCO biosafety laminar flow hood, Synergy HT Microplate reader; UVA-vis Irradiation Equipment; UVA meter; Pipettes
C. Media/Buffers:	Dulbecco's Modified Eagle Medium (DMEM) based medium; Dulbecco's Phosphate-Buffered Saline (DPBS); sterile deionized H_2O
D. Preparation:	Pre-incubate (37°C) tissue inserts in assay medium; Place assay medium and MTT diluent at 4°C, MTT concentrate at -20°C, and record lot numbers of kit components
E. Tissue Culture Plates:	Falcon flat bottom 96-well, 24-well, and 6-well tissue culture plates
F. Reagents:	MTT (*3-4,5-dimethyl thiazole 2-yl*) (1.0mg/mL); Extraction Solution (Isopropanol); Chlorpromazine; Triton X-100 (1%)
G. Other:	Wash bottle; sterile disposable pipette tips; Parafilm; forceps

III. Test Assay

A. Test System

The reconstructed human epidermal model, EpiDerm™ consists of normal human-derived epidermal keratinocytes which have been cultured to form a multilayer, highly differentiated model of the human epidermis. This model consists of organized basal, spinous, and granular layers, and contains a multilayer stratum corneum containing intercellular lamellar lipid layers. The EpiDerm™ tissues are cultured on specially prepared cell culture inserts.

B. Negative Control

Sterile deionized water is used as the negative controls for the EpiDerm™ Phototoxicity assay.

C. Positive Control

Concentrations of chloropromazine, ranging from 0.001% to 0.1%, were used as positive controls for the EpiDerm™ Phototoxicity assay.

D. Data Interpretation Procedure

A photoirritant is predicted if the mean relative tissue viability of the 2 tissues exposed to the test substance and 60 minutes of 6 J/cm^2 is reduced by 20% compared to the non-irradiated control tissues.

IV. Method

A. Tissue Conditioning

Upon MatTek kit arrival at Active Micro Technologies, LLC the tissue inserts are removed from their shipping medium and transferred into fresh media and tissue culture plates and incubated at 37°C at 5% CO_2 and 95% relative humidity for 60 minutes. After those 60 minutes the inserts are transferred into fresh media and tissue culture plates and tissue insert dosing begins.

B. Test Substance Exposure

50µL of the diluted test substance in their respective concentrations are applied to 2 tissue inserts and allowed to incubate for overnight in a humidified incubator (37°C, 5% CO_2, 95% RH).

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C. Tissue Irradiation

Tissue inserts in their 6-well plates are UVA-irradiated for 60 minutes with 6 J/cm^2 at room temperature. The non-irradiated tissue inserts are incubated at room temperature in the dark.

D. Tissue Washing and Post Incubation

After UVA-irradiation and dark incubation is complete the tissue inserts are washed using sterile DPBS and transferred to fresh 6-well plates and media for overnight incubation at 37°C, 5% CO_2, 95% RH.

E. MTT Assay

Tissue inserts are transferred into 300µL MTT media in pre-filled plates and incubated for 3 hours at 37°C, 5% CO_2, and 95% RH. Inserts are then removed from the MTT medium and placed in 2mL of the extraction solution. The plate is sealed and incubated at room temperature in the dark for 24 hours. After extraction is complete the tissue inserts are pierced with forceps and 2 x 200µL aliquots of the blue formazan solution is transferred into a 96 well plate for Optical Density reading. The spectrophotometer reads the 96-well plate using a wavelength of 570 nm.

V. Acceptance Criterion

A. Negative Control

The results of this assay are acceptable if the mean negative control Optical Density (OD_{570}) is ≥ 0.8.

B. Positive Control

The assay meets the acceptance criterion if a dose dependent reduction in cell viability in the UVA-irradiated tissues is between 0.00316% and 0.0316%.

C. Standard Deviation

Since the phototoxicity potential is predicted from the mean viability of 2 tissues for the EpiDerm™ Phototoxicity Protocol, the variability of the replicates should not exceed 30%.

VI. Results

A. Tissue Characteristics

The tissue inserts included in the MatTek EpiDerm™ assay kit were in good condition, intact, and viable.

B. Tissue Viability Assay

The results are summarized in Figure 1. Cell viability is calculated for each tissue as a percentage of the corresponding vehicle control either irradiated or non-irradiated. Tissue viability was not reduced by 20% in the presence of the test substance and UVA-irradiation at concentrations of 0.5%, 1.5%, and 5.0%. The negative control mean exhibited acceptable relative tissue viability while the positive control exhibited dose dependent loss of tissue viability and cell death.

C. Test Validity

The data obtained from this study met criteria for a valid assay. The negative and positive controls performed as anticipated.

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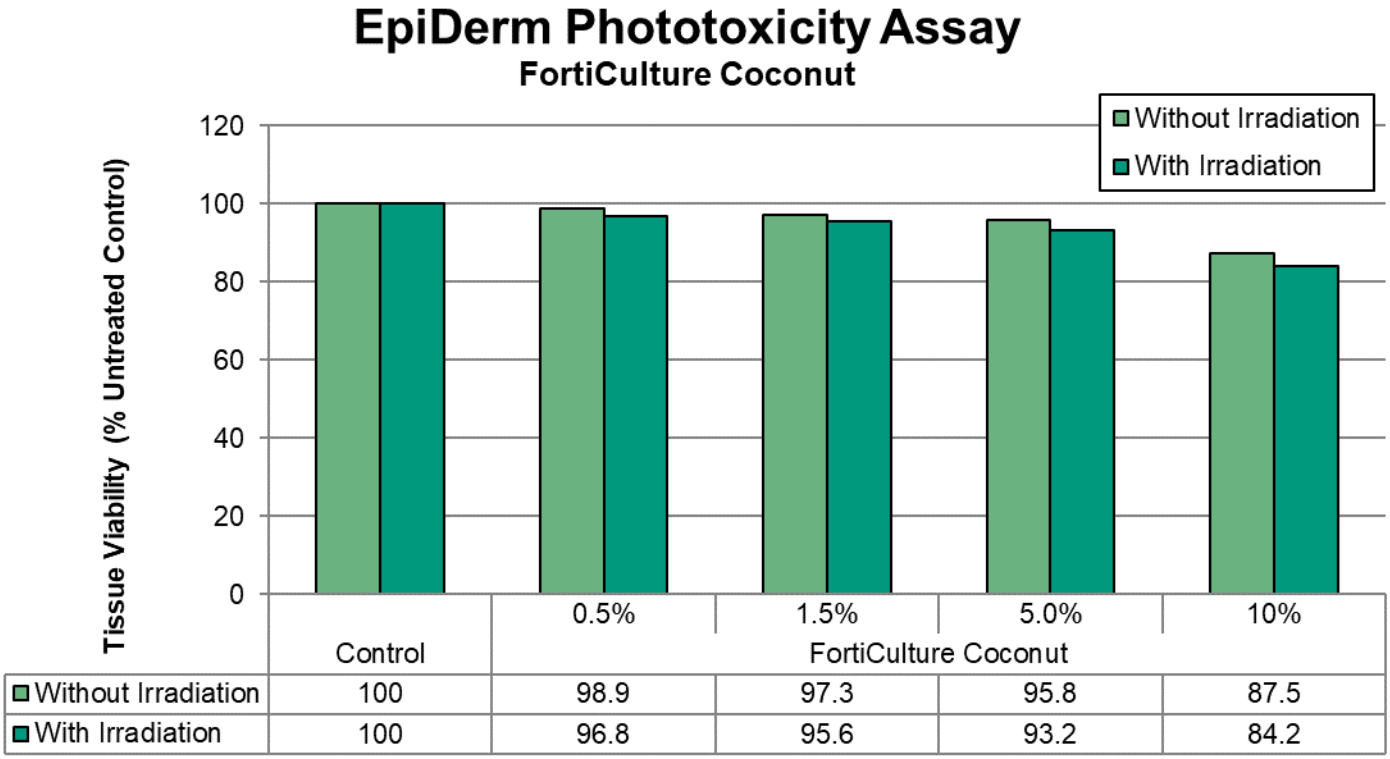
VII. Conclusion

Phototoxicity (photoirritation) is defined as an acute toxic response that is elicited after exposure of the skin to certain chemicals and subsequent exposure to light. Under the conditions of this assay, the test article substance was considered to be **non-phototoxic** at concentrations of 0.5%, 1.5%, 5.0% and 10.0%. The negative and positive controls performed as anticipated

There is a slight decrease in viability at the 10% concentration but viability does not decrease more than the acceptable 20%. We can safely say that **FortiCulture Coconut** is not a photoirritant when used at the suggested use levels of 1.0% - 4.0%.



	Control	0.5%	1.5%	5.0%	10%
Without Irradiation	100	98.9	97.3	95.8	87.5
With Irradiation	100	96.8	95.6	93.2	84.2

Figure 1: EpiDerm Phototoxicity Graph

Certificate of Origin

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FortiCulture Coconut
Code: M11001

Active Micro Technologies, LLC certifies that the above listed ingredient is manufactured in the United States of America.

Active Micro Technologies, LLC certifies that all raw material(s) used to manufacture the above listed ingredient originate in the United States of America. More specifically, the *Cocos nucifera* is sourced from the island of Hawaii.

Active Micro Technologies, LLC certifies that all raw material(s) used to manufacture the above listed ingredient are prepared from non-GMO organisms and are BSE/TSE Free.

Active Micro Technologies, LLC certifies the below sources for each item listed in our INCI Name:

INCI Name	Source
Water	Water
Cocos Nucifera (Coconut) Fruit Extract	Plant (*Cocos nucifera*)
Saccharomyces Ferment Filtrate	Yeast (*Saccharomyces cerevisiae*)

Active Micro Technologies, LLC certifies that the above listed ingredient can be classified as Vegan Compliant.

Active Micro Technologies, LLC certifies that the above listed ingredient has never been tested on animals.

Active Micro Technologies, LLC certifies that the above listed ingredient has the following ISO 16128 value, based on the Compositional Breakdown:

Natural Index (NI)	Natural Origin Index (NOI)
1	1

SECTION 1. IDENTIFICATION

Product Name/Identifier	FortiCulture Coconut
Product Code	M11001
Recommended Use	Ingredients/Raw materials used in the manufacturing of Cosmetic Products'; Antimicrobial
Restrictions on Use	None
Supplier/Manufacturing Site	Active Micro Technologies, LLC
Address	107 Technology Drive Lincolnton, NC 28092, USA
Telephone No. (24hrs)	1-704-276-7100
Fax No.	1-704-276-7101
Emergency Telephone #	1-704-276-7100 (Mon-Fri: 8:00AM – 5:00PM EST)

SECTION 2. HAZARD(S) IDENTIFICATION

Classification:

GHS / CLP
Basis for Classification: Based on present data no classification and labeling is required according to GHS, taking into account the national implementation (United Nations version 2011)

USA
OSHA Regulatory Status: This material is non-hazardous as defined by the American OSHA Hazard Communication Standard (29 CFR 1910.1200).

Europe
Basis for Classification:
-According to present data no classification and labeling is required according to Reg. (EC) No 1272/2008.
-This product is not classified as hazardous to health or environment according to the CLP regulation.

Labeling Elements:

Pictograph: No hazard symbol expected

Hazard statements/Signal Word: Not applicable

Precautionary statements:
P233: Keep container tightly closed
P281: Use personal protective equipment as required
P402: Store in a dry place
P404: Store in a closed container
P410: Protect from sunlight
P411: Store at temperatures not exceeding 32°C

Other hazards which do not result in classification:

No particular fire or explosion hazard.
By mechanical effect: No particular hazards.
By hygroscopic effect: No particular hazards.

US NFPA 704 (National Fire Protection Association) Hazard Rating System:

Health hazard: Rating 0; Normal Material
Flammability: Rating 0, Will Not Burn
Reactivity: Rating 0, Stable
Other Hazard Information: None

Results of PBT and vPvB assessment:

-PBT: Not applicable
-vPvB: Not applicable

SECTION 3. COMPOSITION / INFORMATION ON INGREDIENTS

Common Chemical Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate

Generic name:

Chemical Family: Ferment

Description: Mixture: consisting of the following components. This section describes all components of the mixture

Substance	CAS Numbers	EC Numbers	Percentage
Water	7732-18-5	231-791-2	80.00%
Cocos Nucifera (Coconut) Fruit Extract	8001-31-8	232-282-8	10.00%
Saccharomyces Ferment Filtrate	8013-01-2	232-387-9	10.00%

Formula: Not applicable

SECTION 4. FIRST-AID MEASURES

General: In all cases of doubt, or when symptoms persist, seek medical attention.

Inhalation: Move to fresh air from exposure area. Get medical attention for any breathing difficulty.

Skin contact: Rinse with soap and water. Get medical advice if irritation develops.

Eye contact:	Immediately rinse with water for at least 15 minutes, while keeping the eyes wide open. Consult with a physician.
Ingestion:	Consult with a physician.
Protection of first-aiders:	No special protection required.

SECTION 5. FIRE-FIGHTING MEASURES

Fire and explosion hazards: Not considered to be a fire and explosion hazard

Extinguishing media:

Suitable:	Water, dry chemicals, foam & carbon dioxide.
Not suitable:	None known

Fire fighting: Move container from fire area if it can be done without risk.
Avoid inhalation of material or combustion by-products.
Stay upwind and keep out of low area

Protection for fire-fighters: Boots, gloves, goggles.

SECTION 6. ACCIDENTAL RELEASE MEASURES

Personal precautions: Avoid contact with eyes.
Personal Protective Equipment:
-Protective goggles

Environmental precautions: Prevent entry into sewers and waterways. Do not allow material to contaminate ground water system

Methods for cleaning up:

Recovery:	Pick up free liquid for recycling or disposal. Residual liquid can be absorbed on an inert material. Sweep or vacuum up any powder and place in a clearly labeled waste container, avoiding dust formation.
Cleaning/Decontamination:	Wash non-recoverable remainder with water.
Disposal:	For disposal of residues refer to sections 8 & 13.

SECTION 7. HANDLING AND STORAGE

Handling

Technical measures:	Labeling: Keep out of the reach of children.
Measures:	For industrial use, only as directed.
Safe handling advice:	Wash hands after use. Avoid storage near feed or food stuff.

Storage

Technical measures:	Keep container closed.
Recommended Storage Conditions:	Store in a dry place at temperatures not exceeding 32°C. Based on stability studies, the optimum storage temperature for maximization of shelf life is 23 - 25°C. However, it may be stored at temperatures between 16 and 32°C if such specific temperature control is not available. Do not freeze. Please refer to stability data for effects heat or cold may have on the specifications of the product.
Incompatible products:	Avoid contact with strong oxidizers. Refer to the detailed list of incompatible materials (Section 10 Stability/Reactivity)
Packaging:	Product may be packaged in normal commercial packaging.
Packaging materials:	Recommended - Polypropylene & High Density Polyethylene

SECTION 8. EXPOSURE CONTROLS / PERSONAL PROTECTION

Precautionary statements: Ensure adequate ventilation

Control parameters

Occupational exposure Limits:

France:	Not Determined
ACGIH:	Not Determined
Korea:	Not Determined
UK:	Not Determined
Surveillance procedures:	Not Determined
Engineering measures:	Not Determined

Personal Protective Equipment:

Respiratory protection:	Local exhaust
Hand protection:	Protective gloves made of rubber or neoprene.
Eye protection:	Safety glasses.
Collective emergency equipment:	Eye fountain.
Skin and Body Protection:	Suitable protective clothing
Hygiene measures:	Handle in accordance with good industrial hygiene and safety practice.
Measures related to the Environment:	No particular measures.

SECTION 9. PHYSICAL AND CHEMICAL PROPERTIES

Appearance:	Clear to slightly hazy liquid
Color:	Colorless to slightly light yellow
Odor:	Characteristic

pH (Direct):	3.8 – 5.5
NVM (1g-1hr-105°C):	16.0 – 22.0%
Lipopeptide Content (Kjeldahl Nitrogen x 6.25):	15.0 – 20.0%
Keratinocyte Expressed Proteins (KEP) > 5%:	Positive
Heavy Metals:	< 20 ppm
Lead:	< 10 ppm
Arsenic:	< 2 ppm
Cadmium:	< 1 ppm
Minimum Inhibitory Concentration Organism (ATCC#):	
E. coli (#8739):	0.50 – 2.00%
S. aureus (#6538):	1.00 – 2.00%
P. aeruginosa (#9027):	0.25 – 1.00%
C. albicans (#10231):	1.00 – 2.00%
A. brasiliensis (#16404):	1.00 – 2.00%
Specific Gravity:	Not determined
Vapor density:	Not applicable
Boiling Point:	100°C
Freezing Point:	0°C
Melting point:	Not applicable
Flash point:	> 93°C
Oxidizing properties:	Non oxidizing material according to EC criteria.
Solubility:	
In water:	Soluble
In organic solvents:	Not determined
Log P:	Not determined

SECTION 10. STABILITY AND REACTIVITY

Stability:	Stable under ordinary conditions of use and storage up to one year then re-test to full product specifications to extend shelf life
Hazardous reactions:	None known
Conditions to avoid:	No dangerous reactions known under use of normal conditions. Avoid extreme heat.

Materials to avoid: No dangerous reaction known with common products.

Hazardous decomposition products: None known

SECTION 11. TOXICOLOGICAL INFORMATION

Ingestion: Not Determined
Dermal: Non-Irritant (Dermal Irritection Model)
Ocular: Non-Irritant (Ocular Irritection Model)
Inhalation: Not Determined

Acute toxicity data: Non-Irritant, Non-Primary Sensitizer & Non-Photo Irritant

Sensitization: Non-Primary Irritant & Non-Primary Sensitizers; Will not cause allergic contact dermatitis (In Chemico Skin Sensitization Direct Peptide Reactivity Assay & In Vitro Skin Sensitization ARE-Nrf2 Luciferase Test Method)

Repeated dose toxicity: No known effects
Subacute to chronic toxicity: Not Determined

Mutagenicity: Non-Mutagenic (OECD471/ISO10993.Part 3 – Genotoxicity: Bacterial Reverse Mutation Test)

Additional Toxicological Information: This product is not subject to classification according to the calculation method of the General EU Classification Guidelines for Preparations as issued in the latest version.

Specific effects:
Carcinogenicity: No known effects
Mutagenicity: No known effects
Reproductive toxicity: No known effects
Neuro-toxicity: No known effects

For more information: Does not present any particular risk on handling under normal conditions of good occupational hygiene practice.

This product has not been tested for the following:
-Primary cutaneous and corrosive irritation
-Acute oral toxicity
-Mutagenicity/genotoxicity

SECTION 12. ECOLOGICAL INFORMATION

Ecotoxicity
Effects on the aquatic environment: EC_{10} (Freshwater Alga): 103.68 mg/L - Not harmful to aquatic organisms
EC_{20} (Freshwater Alga): 184.24 mg/L - Not harmful to aquatic organisms

Biodegradability:
Persistence: Readily Biodegradable (94.2% biodegradation after 28 days of testing)

Bioaccumulation:
Octanol / water partition coefficient: Not Determined

Mobility:
Precipitation:
Expected behavior of the product: Ultimate destination of the product: Soil & sediment.

Other Adverse Effects: None known

SECTION 13. DISPOSAL CONSIDERATIONS

Residues from product

Prohibition: Do not allow the product to be released into the Environment.
Destruction/Disposal: Dispose of in accordance with relevant local regulations

Contaminated packaging

Decontamination/cleaning: Cleaning is not required prior to disposal.
Destruction/Disposal:

Note: Take all necessary precautions when disposing of this product according to local regulations.

SECTION 14. TRANSPORT INFORMATION

UN Number: None
UN Shipping Name: None
Transport Hazard Class: Not classified as dangerous for transport

Land (rail/road): Material is not restrictive for land transport and is not regulated by ADR/RID
Sea: Material is not restrictive for sea transport and is not regulated by IMO/IMDG
Air: Material is not restrictive for air transport and is not regulated by ICAO/IATA

Marine Pollutant: No

Transport/Additional Information: Not regulated for US DOT Transport in non-bulk containers
This material is not dangerous or hazardous

Special Precautions for User: None known

The above regulatory prescriptions are those valid on the date of publication of this sheet. However, given the possible evolution of transport regulations for hazardous materials and in the event of the MSDS in your possession dating back more than 12 months, it is advisable to check their validity with your sales office.

SECTION 15. REGULATORY INFORMATION

Labeling:

EC regulations: This product does not need to be labeled in accordance with EC Directives or respective national laws

Further regulations

United Kingdom: Handle in accordance with relevant British regulation: control of substance Hazardous to Health Regulations Environmental Hygiene Guidance: EH40
Workplace Exposure Limits (revised annually)

Korea regulations:

Industrial safety and hygiene regulation:	No
Hazardous material control regulation:	No
Fire prevention regulation:	No

Other regulations:

EINECS inventory status:	Aqua:	231-791-2
	Cocos Nucifera Fruit Extract:	232-282-8
	Saccharomyces Ferment Filtrate:	232-387-9
TSCA inventory status:	Exempt	
AICS inventory status:	7732-18-5 & 8001-31-8 & 8013-01-2	
Canadian (CEPA DSL) inventory status:	Listed as Water (DSL) & Coconut Oil (DSL) & Yeast, ext. (DSL)	
Japan (MITI list):	Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate	
Korea:	Water & Cocos Nucifera (Coconut) Fruit Extract^ & Saccharomyces Ferment Filtrate	
China inventory status:	Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate	
Philippines inventory status:	Listed as Water & Coconut Oil & Yeast, ext.	

^Not listed in 2004 CTFA Dictionary – Registered with Personal Care Products Council

Note: The regulatory information given above only indicates the principal regulations specifically applicable to the products described in this sheet. The user's attention is drawn to the possible existence of additional provision which complete these regulations. Please refer to all applicable international, national and local regulations and provisions

SECTION 16. OTHER INFORMATION

Prohibited uses: For specific uses, food industry, ask the manufacturer for more information.

Last Revision Date: 08/16/2023

Preparation Date: 10/12/2023

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MSDS summary of changes - Updated NVM – Section 9 (Physical & Chemical Properties)

The information given is based on our knowledge of this product, at the time of publication in good faith. The attention of the user is drawn to the possible risks incurred by using the product for any other purpose other than which it was intended. This is not in any way excuse the user from knowing and applying all the regulations governing their activity. It is sole responsibility of the user to take all precautions required in handling the product. The purpose of mandatory regulation mentioned is to help the user to fulfill his obligations regarding the use of products. This information is not exhaustive, this is not exonerate the user from ensuring that legal obligations other than those mentioned, relating to the use and storage.

M11001-FortiCulture Coconut Manufacturing Flow Chart

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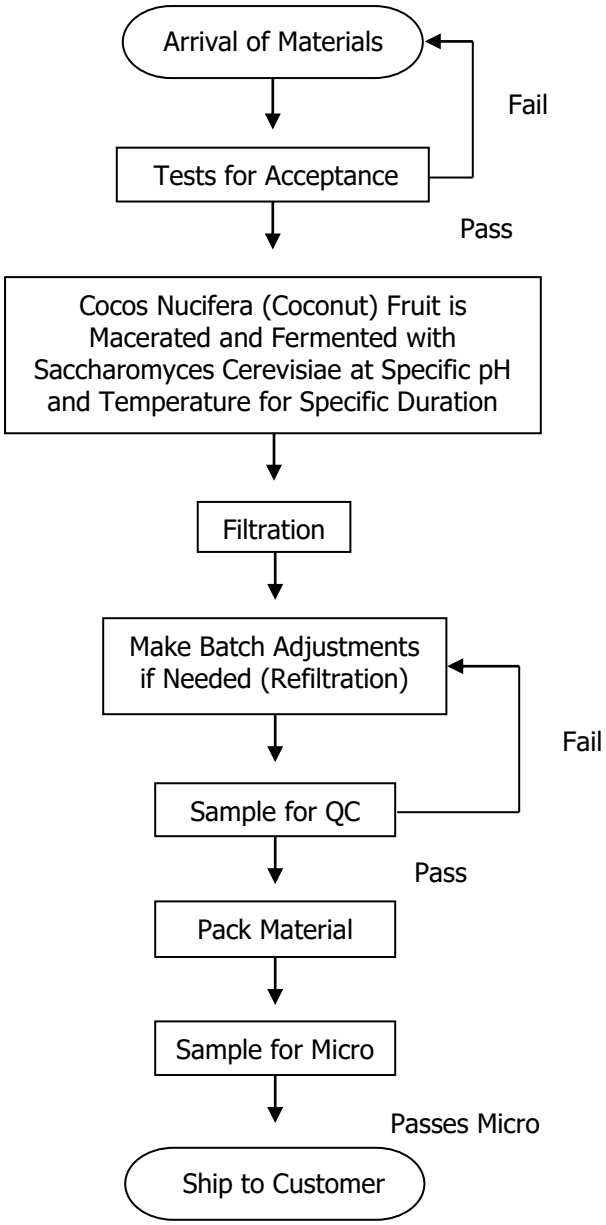

Version#2/08-28-23

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FortiCulture Coconut
Certificate of Compliance

Code: M11001
INCI Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate
INCI Status: Conforms
CAS #: 7732-18-5 & 8001-31-8 & 8013-01-2
EINECS #: 231-791-2 & 232-282-8 & 232-387-9
China NMPA #: N/A

The following information on regulatory clearances is believed to be accurate and is given in good faith as a guide to a global use of our ingredients in cosmetic applications. No representation or warranty as to its competences or accuracy is made. Information is offered for use in general cosmetic applications and may vary in particular applications. Users are responsible for determining the suitability of these products for their own particular use. All regulatory decisions should be made on the advice of your regulatory group or legal counsel.

Country / Regulatory Body	Status of Product
EU (CosIng)	Compliant
USA (TSCA)	Compliant
Australia (AICS)	Compliant
Japan (METI)	Compliant
Canada (DSL)	Compliant
China (IECIC)	Compliant
Brazil (ANVISA)	Compliant
Korea (KECI)	Compliant
Philippines (PICCS)	Compliant
Mexico (COFEPRIS)	Compliant

Version#1/07-11-23/Form#62

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FortiCulture Coconut
Code: M11001

Attention must be paid to the use of FortiCulture Coconut in the equivalent of OTC formulations (eg. quasi-drugs in Japan, or therapeutic goods in Australia). Some countries maintain restricted inventories of raw materials that can be used in those applications so more detailed guidance may be required.

FortiCulture Coconut and any components or impurities are in compliance with the rules governing cosmetic products in the European Union (Directive 76/768/ECC & Regulation No. 1223/2009). The recommended use levels for FortiCulture Coconut is 1.00 – 4.00%.

FortiCulture Coconut is in compliance with the standardized set of rules developed and approved by the NPA (Natural Products Association).

The Nagoya Protocol provides a scheme for the fair and equitable sharing of benefits derived from Genetic Resources. Information regarding the Nagoya Protocol and Access and Benefit Sharing (ABS) is available at https://www.cbd.int/abs/. The agreement focusses on wild taxa and excludes most commercially cultivated crops. For the signatories to the agreement, responsibility for Benefit Sharing falls on the entity exporting or extracting the resource from the signatory country. Active Micro Technologies audits its suppliers to conform compliance with the Nagoya Protocol where applicable.

FortiCulture Coconut is considered a non-hazardous material. All significant toxicological routes of absorption have been considered as well as the systemic effects and margin of safety (MoS) based on a no observed adverse effects level (NOAEL). Due to the restriction placed on animal testing of cosmetic raw materials, and Active Micro Technologies, LLC's internal non-animal testing policy, this product was not tested for NOAEL.

FortiCulture Coconut was tested using *in vitro* dermal and ocular irritation models. This product was found to be non-irritating in both models.

To our knowledge the above material is free of CMR (*) substances, as defined according to Regulation (EC) No 1272/2008 and Cosmetic Regulation (EC) No 1223/2009 as amended. Products supported for Personal Care applications will not be classified as CMR (*), as defined by (EC) 1272/2008 on the Classification, Labelling and Packaging of Substances and Mixtures, unless supported by a positive SCCS opinion.

(*) Carcinogenic, Mutagenic, toxic for Reproduction

Active Micro Technologies, LLC certifies that to the best of our knowledge our product does not contain any material listed on California Proposition 65.

As of July 11, 2023, FortiCulture Coconut does not contain any substances present on the so called "candidate list" provided by the European Chemicals Agency (ECHA). We further certify that this material has not been manufactured using any of the species listed in the CITES Appendices as of July 11, 2023.

FortiCulture Coconut is REACH Compliant and free of the following:

- Butylphenyl methylpropional (Lilial)
- Formaldehyde or formaldehyde donors
- Glycol ethers
- Gluten
- Lactose
- Nanoparticles
- Nitrosamines
- Palm oil/palm kernel oil (or derivatives)
- Parabens
- Paraffin/petroleum products
- Phthalates
- Polyethylene glycol (PEG)
- Residual solvents
- Sulfates
- Volatile organic compounds

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Product Name: FortiCulture Coconut
Code: M11001
INCI Name: Water & Cocos Nucifera (Coconut) Fruit Extract & Saccharomyces Ferment Filtrate

FortiCulture Coconut is manufactured by fermenting macerated *Cocos nucifera* (coconut) fruit with *Saccharomyces cerevisiae* at a specific pH and temperature for a set duration. Lastly, the entire mixture is filtered.

FortiCulture Coconut was tested using *in vitro* dermal and ocular irritation models, including phototoxicity irritation (EpiDerm™ EPI-200-SIT). This product was found to be non-irritating in all models, including non-phototoxic for the *in vitro* dermal model.

A *Salmonella typhimurium* reverse mutation standard plate incorporation study was conducted to evaluate whether FortiCulture Coconut would cause mutagenic changes in the average number of revertants for histidine-dependent *Salmonella typhimurium* strains in the presence and absence of S9 metabolic activation. This study was conducted to satisfy, in part, the Genotoxicity requirement of the International Organization for Standardization: Biological Evaluation of Medical Devices, Part 3: Tests for Genotoxicity, Carcinogenicity and Reproductive Toxicity. FortiCulture Coconut was considered to be non-mutagenic to the *Salmonella typhimurium* tester strains under the conditions of this assay.

FortiCulture Coconut was also tested via the OECD TG 442C Direct Peptide Reactivity (DPRA) and OECD TG 442D In Vitro Skin Sensitization (KeratinoSens™) Assays in accordance with the EURL ECVAM and UN GHS guidelines. This product was determined to be a non-skin sensitizer in both *in chemico* and *in vitro* models. Additionally, OECD TG 497 (Defined Approaches on Skin Sensitization) outlines the '2 out of 3' direct approach to predict a skin sensitization hazard. Specifically, if assays for two of the first three key events in the adverse outcome pathway leading to skin sensitization provide consistent results, then the substance is predicted accordingly as a sensitizer or non-sensitizer. In line with OECD TG 497, FortiCulture Coconut is predicted as a non-sensitizer given the concordant results obtained from our DPRA and KeratinoSens™ studies, therefore OECD TG 442E (h-CLAT) is not required.

A Freshwater Alga Growth Inhibition test via OECD 201 was subsequently performed to determine the potential toxicity of FortiCulture Coconut. In this assay, *Psuedokirchneriella subcatpitata* are exposed to the test substance for 72 hours and growth and growth inhibition through cell count against a control is performed. The response is evaluated as a function of the exposure concentration in comparison with the average growth of replicate, unexposed control cultures. After 72 hours, the percent inhibition for FortiCulture Coconut was determined to be 103.68 mg/L EC_{10} and 184.24 mg/L EC_{20}. The results of this assay indicate that the product is not classified and therefore not harmful to aquatic organisms.

FortiCulture Coconut was also assessed for ready biodegradability in an aerobic aqueous medium via the OECD 301 B Ready Biodegradability: CO_2 Evolution (Modified Sturm Test). FortiCulture Coconut achieved 94.2% biodegradation after 28 days of testing, indicating that the product meets method requirements for the Ready Biodegradable classifications.

The full reports for each safety study analyzing FortiCulture Coconut are attached for reference.

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Due to the restriction placed on animal testing of cosmetic raw materials, and Active Concepts, LLC's internal non-animal testing policy, FortiCulture Coconut was not tested for NOAEL. However, NOAEL values for all INCI ingredients have been established and are summarized in Table 1. Provided NOAEL values and other safety and toxicity studies are available for *Cocos nucifera* (coconut) Fruit Extract and *Saccharomyces cerevisiae* Ferment Filtrate, it is estimated that FortiCulture Coconut will have a NOAEL of at least 330 mg/kg/day.[1-11]

Table 1. FortiCulture Coconut INCI Ingredients and established NOAEL values.

INCI Ingredient	NOAEL Value (mg/kg/day)	Reference
Water	-	-
Cocos nucifera (coconut) Fruit Extract	330	5-7
Saccharomyces cerevisiae Ferment Filtrate	800	8

Cocos nucifera (coconut) fruit is regularly consumed as food and daily exposure from ingestion would result in much larger systemic exposures compared to use in topical cosmetic products, and despite an assumed NOAEL greater than 1,000 mg/kg/day, sub-chronic or chronic *in vivo* toxicity studies are not expected to be published.[1-4] Without sub-chronic or chronic toxicity studies available for *Cocos nucifera* (coconut) fruit extracts, an uncertainty factor can be implemented to scientifically derive toxicity from repeated-dose studies (28 days) to sub-chronic toxicity (90 days).[5,6] In accordance with the SCCS Notes of Guidance and supporting literature, applying an uncertainty factor of 3 to the 28-day repeated-dose study administering a *Cocos nucifera* (coconut) fruit extract at 995 mg/kg/day enables a NOAEL of 331 mg/kg/day to be extrapolated for *Cocos nucifera* (coconut) fruit extracts.[5-7]

FortiCulture Coconut is typically comprised of 80.0% water, 10.0% *Cocos nucifera* (coconut) Fruit Extract, and 10.0% *Saccharomyces cerevisiae* Ferment Filtrate. Table 2 demonstrates 1) the amount of FortiCulture Coconut's INCI ingredients in a final end-product if utilized at the maximum recommended use level (4.0%), 2) the maximum daily exposure of FortiCulture Coconut's INCI ingredients if 18.0 g of a final-end product are applied and assuming 100% absorption, and 3) the established NOAELs for each respective INCI ingredient.

For example, if a consumer (60 kg) applies 18.0 g of a final end-product with 4.0% FortiCulture Coconut every day, a daily exposure of 1.2 mg/kg/day is expected for *Cocos nucifera* (coconut) Fruit Extract (18.0 g x 4.0% use-level x 10.0% composition = 72 mg; 72 mg / 60 kg). Importantly, all of the maximum daily exposure values in Table 2 are well below their respective established NOAEL values.[5-8]

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Table 2. FortiCulture Coconut INCI Ingredients, in a Final End-Product at 4.0%, Maximum Daily Exposure with 18.0 g Application, and NOAEL Values.

INCI Ingredient	Compositional Breakdown (%)	Amount in Final End-Product at 4.0% (%)	Maximum Daily Exposure, 18.0 g Application, 100% Absorption Assumption (mg/kg/day)	NOAEL (mg/kg/day)
Water	80.0	3.2	9.6	-
Cocos nucifera (coconut) Fruit Extract	10.0	0.4	1.2	330[5-7]
Saccharomyces cerevisiae Ferment Filtrate	10.0	0.4	1.2	800[8]

Published toxicity and safety studies for INCI ingredients in FortiCulture Coconut are summarized in Table 3.

Table 3. Published Toxicity and Safety Studies for INCI Ingredients in FortiCulture Coconut.

INCI Ingredient	Toxicity / Safety Studies Conducted	Results	Reference
Cocos nucifera (coconut) Fruit Extract	Acute, repeated-dose, reproductive, & developmental toxicity, genotoxicity, mutagenicity, dermal & ocular irritation & sensitization, oral consumption by humans	Non-toxic, non-genotoxic, non-mutagenic, non-irritating, non-sensitizing, well-tolerated by humans	1-4, 7
Saccharomyces cerevisiae Ferment Filtrate	Acute, sub-chronic, & chronic toxicity, genotoxicity, mutagenicity, cytotoxicity, dermal & ocular irritation & sensitization	Non-toxic, non-genotoxic, non-mutagenic, non-cytotoxic, non-irritating, non-sensitizing	8, 9

Additionally, *Cocos nucifera* (coconut) and *Saccharomyces cerevisiae* are commonly used in the food and nutraceutical industries. Since *Cocos nucifera* (coconut) and *Saccharomyces cerevisiae* are intentionally used in food, their extracts/ferments may be classified as Generally Recognized as Safe (GRAS) according to the FDA's Federal Food, Drug and Cosmetic Act of 1938 (FD&C Act) and Modernization of Cosmetics Regulation Act of 2022 (MoCRA).[10,11]

The act states:

> Any substance that is intentionally added to food is a food additive, that is subject to premarket review and approval by FDA, unless the substance is generally recognized, among qualified experts, as having been adequately shown to be safe under the conditions of its intended use, or unless the use of the substance is otherwise excluded from the definition of a food additive.[10,11]

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The Cosmetic Ingredient Review (CIR) also published reports assessing the safety of *Cocos nucifera* (coconut) Fruit Extract and *Saccharomyces cerevisiae* Ferment Filtrate. The CIR reports concluded that *Cocos nucifera* (coconut) Fruit Extract and *Saccharomyces cerevisiae* Ferment Filtrate are safe for use in cosmetic formulations in the present practices of use and concentration.[2-4,9]

Several published data sets exist to support the safety of FortiCulture Coconut. It is presented in an aqueous carrier, all but eliminating its risk for inhalation. Toxicological, irritation, and sensitization assays have all been performed with favorable results for each. This knowledge combined with the tested and published toxicity assays allows us to support the safety of FortiCulture Coconut in cosmetic applications.

It is logically concluded that FortiCulture Coconut is safe in cosmetic applications at use levels of 1.0 – 4.0%. No further testing is required at this time.

References

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2. Cosmetic Ingredient Review. Safety Assessment of Cocos nucifera (Coconut)-Derived Ingredients as Used in Cosmetics. Online Access: https://www.cir-safety.org/ingredients
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10. U.S. Food and Drug Administration (FDA). Federal Food, Drug, and Cosmetic Act of 1938 (FD&C Act). Online Access: www.fda.gov
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